HUMANIZED IMMUNOGLOBULINS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation of U.S.S.N. 08/484,537 filed June 7, 1995, which is a divisional of U.S.S.N. 07/634,278 filed December 19, 1990, now U.S. Patent No. 5,530,101, which is a continuation-in-part of U.S.S.N. 07/590,274 filed September 28, 1990 and of U.S.S.N. 07/310,252 filed February 13, 1989, which is a continuation-in-part of U.S.S.N. 07/290,975 filed December 28, 1988, the disclosures of which are incorporated by reference.

Field of the Invention

The present invention relates generally to the combination of recombinant DNA and monoclonal antibody technologies for developing novel therapeutic agents and, more particularly, to the production of non-immunogenic antibodies having strong affinity for a predetermined antigen.

Background of the Invention

The advent of monoclonal antibody technology in the mid 1970's heralded a new age of medicine. For the first time, researchers and clinicians had access to essentially unlimited quantities of uniform antibodies capable of binding to a predetermined antigenic site and having various immunological effector functions. These proteins, known as "monoclonal antibodies" were thought to hold great promise in, e.g., the removal of harmful cells in vivo. Indeed, the clinical value of monoclonal antibodies seemed limitless for this use alone.

Unfortunately, the development of appropriate therapeutic products based on these proteins has been severely hampered by a number of drawbacks inherent in monoclonal antibody production. For example, most monoclonal antibodies are mouse derived, and thus do not fix human complement well. They also lack other important immunoglobulin functional characteristics when used in humans.

Perhaps most importantly, non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that after injection of a foreign antibody, the immune response mounted by a patient can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after one or several treatments with any non-human antibodies, subsequent treatments, even for unrelated therapies, can be ineffective or even dangerous in themselves, because of cross-reactivity.

While the production of so called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. Moreover, efforts

to immortalize human B-cells or generate human hybridomas capable of producing human immunoglobulins against a desired antigen have been generally unsuccessful, particularly with many important human antigens. Most recently, recombinant DNA technology has been utilized to produce immunoglobulins which have human framework regions combined with complementarity determining regions (CDR's) from a donor mouse or rat immunoglobulin (see, e.g., EPO Publication No. 0239400, which is incorporated herein by reference). These new proteins are called "reshaped" or "humanized" immunoglobulins and the process by which the donor immunoglobulin is converted into a human-like immunoglobulin by combining its CDR's with a human framework is called "humanization". Humanized antibodies are important because they bind to the same antigen as the original antibodies, but are less immunogenic when injected into humans.

However, a major problem with present humanization procedures has been a loss of affinity for the antigen (Jones et al., Nature, 321, 522-525 (1986)), in some instances as much as 10-fold or more, especially when the antigen is a protein (Verhoeyen et al., Science, 239, 1534-1536 (1988)). Loss of any affinity is, of course, highly undesirable. At the least, it means that more of the humanized antibody will have to be injected into the patient, at higher cost and greater risk of adverse effects. Even more critically, an antibody with reduced affinity may have poorer biological functions, such as complement lysis, antibody-dependent cellular cytotoxicity, or virus neutralization. For example, the loss of affinity in the partially humanized antibody HuVHCAMP may have caused it to lose all ability to mediate complement lysis (see, Riechmann et al., Nature, 332, 323-327 (1988); Table 1).

Thus, there is a need for improved means for producing humanized antibodies specifically reactive with strong affinity to a predetermined antigen. These humanized immunoglobulins should remain substantially non-immunogenic in humans, yet be easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

Summary of the Invention

The present invention provides novel methods for preparing humanized immunoglobulin chains having generally one or more complementarity determining regions (CDR's) from a donor immunoglobulin and a framework region from a human immunoglobulin. The preferred methods comprise first comparing the framework or variable region amino acid sequence of the donor immunoglobulin to corresponding sequences in a collection of human immunoglobulin chains, and selecting as the human immunoglobulin one of the more homologous sequences from the collection. The human immunoglobulin, or acceptor immunoglobulin, sequence is typically selected from a collection of at least 10 to 20 immunoglobulin variable region sequences, and usually will have the highest homology to the donor immunoglobulin sequence of any sequence in the collection. The human immunoglobulin framework sequence will typically have about 65 to 70% homology or more to the donor

immunoglobulin framework sequences. The donor immunoglobulin may be either a heavy chain or light chain, and the human collection will contain the same kind of chain. A humanized light and heavy chain can be used to form a complete humanized immunoglobulin or antibody, having two light/heavy chain pairs, with or without partial or full-length human constant regions.

To form the humanized variable region, amino acids in the human acceptor sequence will be replaced by the corresponding amino acids from the donor sequence if they are in the category

(1) the amino acid is in a CDR.

In another embodiment of the present invention, either in conjunction with the above comparison step or separately, additional amino acids in the acceptor immunoglobulin chain may be replaced with amino acids from the CDR-donor immunoglobulin chain. More specifically, further optional substitutions of a human framework amino acid of the acceptor immunoglobulin with the corresponding amino acid from a donor immunoglobulin will be made at positions which fall in one or more of the following categories:

(2) the amino acid in the human framework region of the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is common for that position in human immunoglobulin sequences; or

(3) the amino acid is immediately adjacent to one of the CDR's; or

(4) the amino acid is predicted to be within about 3A of the CDR's in a three-dimensional immunoglobulin model and capable of interacting with the antigen or with the CDR's of the donor or humanized immunoglobulin.

Moreover, an amino acid in the acceptor sequence may optionally be replaced with an amino acid typical for human sequences at that position if

(5) the amino acid in the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is also rare, relative to other human sequences.

The humanized immunoglobulin chain will typically comprise at least about 3 amino acids from the donor immunoglobulin in addition to the CDR's, usually at least one of which is immediately adjacent to a CDR in the donor immunoglobulin. The heavy and light chains may each be designed by using any one or all three of the position criteria.

When combined into an intact antibody, the humanized light and heavy chains of the present invention will be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen (such as a protein or other compound containing an epitope). These affinity levels can vary from about 10^8 M^{-1} or higher, and may be within about 4 fold, preferably within about 2 fold of the donor immunoglobulin. Ideally, the humanized antibodies will exhibit affinity levels at least about 60 to 90% of the donor immunoglobulin's original affinity to the antigen.

Once designed, the immunoglobulins, including binding fragments and other immunoglobulin forms, of the present invention may be produced

readily by a variety of recombinant DNA or other techniques. Preferably, polynucleotides encoding the desired amino acid sequences are produced synthetically and by joining appropriate nucleic acid sequences, with ultimate expression in transfected cells. Notably, the methods of the present invention maximize the likelihood of producing humanized immunoglobulins with optimum binding characteristics without the need for producing intermediate forms that may display stepwise improvements in binding affinity. The humanized immunoglobulins will be particularly useful in treating human disorders susceptible to monoclonal antibody therapy, but find a variety of other uses as well.

BRIEF DESCRIPTION OF THE FIGURES

Fig. 1A and Fig. 1B. Amino acid sequences (1-letter code) of the light chain (A) (SEQ ID NOS:1 and 2) and heavy chain (B) (SEQ ID NOS:3 and 4) variable regions of the mouse anti-Tac antibody (upper lines), compared with the human Eu antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the Eu antibody framework replaced with mouse amino acids in the humanized antibody are double underlined. The number of the first position on each line is given on the left.

Fig. 2A and Fig. 2B. Amino acid sequences (1-letter code) of the light chain (A) (SEQ ID NOS:46 and 47) and heavy chain (B) (SEQ ID NOS:48 AND 49) variable regions of the mouse Fd79 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

Fig. 3A and Fig. 3B. Amino acid sequences (1-letter code) of the light chain (A) (SEQ ID NOS:50 and 51) and heavy chain (B) (SEQ ID NOS:52 and 53) variable regions of the mouse Fd138-80 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

Fig. 4A and Fig. 4B. Amino acid sequences (1-letter code) of the light chain (A) (SEQ ID NOS:54 and 55) and heavy chain (B) (SEQ ID NOS:56 and 57) variable regions of the mouse M195 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

Fig. 5A and Fig. 5B. Amino acid sequences (1-letter code) of the light chain (A) (SEQ ID NOS:58 and 59) and heavy chain (B) (SEQ ID NOS:60 and 61) variable regions of the mouse mik-β1 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

Fig. 6A and Fig. 6B. Amino acid sequences (1-letter code) of the light chain (A) (SEQ ID NOS:62 and 63) and heavy chain (B)(SEQ ID NOS:64 and 65) variable regions of the mouse CMV5 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

Fig. 7A through Fig. 7D. Fluorocytometry of HUT-102 and Jurkat cells stained with anti-Tac antibody or humanized anti-Tac antibody followed respectively by fluorescein-conjugated goat anti-mouse Ig antibody or goat anti-human Ig antibody, as labeled. In each panel, the dotted curve shows the results when the first antibody was omitted, and the solid curve the results when the first and second (conjugated) antibodies were included as described.

Fig. 8A and Fig. 8B. (A) Fluorocytometry of HUT-102 cells stained with 0-40 ng of anti-Tac as indicated, then with biotinylated anti-Tac, and then with phycoerythrin-conjugated avidin. (B) Fluorocytometry of HUT-102 cells stained with the indicated antibody, then with biotinylated anti-Tac, and then with phycoerythrin-conjugated avidin.

Fig. 9A and Fig. 9B. Schematic diagram of the plasmids pVg1 (A) and pVk (B). The plasmid pVg1 was constructed from the following fragments: an approximately 4850 base pair BamHI-EcoRI fragment from the plasmid pSV2hph containing the amp and hyg genes; a 630-bp fragment containing the human cytomegalovirus IE1 gene promoter and enhancer flanked at the 5' and 3' by EcoR1 and Xba1 linkers respectively; and a 2800 bp XbaI-BamHI fragment containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 gene and the gpt replacing the hyg gene.

Fig. 10A and Fig. 10B. Amino acid sequences of the heavy (A) (SEQ ID NOS:5 and 6) and light (B) (SEQ ID NOS:7 and 8) chain variable regions of the PDL and CDR-only humanized anti-Tac antibodies. The PDL sequence is

shown on the upper line, the CDR-only sequence below. Amino acid differences are boxed. Complementarity Determining Regions (CDR's) are underlined.

Fig. 11A and Fig. 11B. Double-stranded DNA sequence of fragments encoding the heavy (A) (SEQ ID NO:9) and light (B) (SEQ ID NO:10) chain variable regions of the CDR-only humanized anti-Tac antibody including signal sequences. Oligonucleotides used for gene synthesis are marked by solid lines: above, for oligonucleotides from upper strand, and below, for oligonucleotides from lower strand. Restriction sites used for cloning are underlined.

Fig. 12. FACS analysis of HUT-102 cells stained with PDL and CDR-only humanized anti-Tac antibodies and negative control antibody Fd79.

Fig. 13. Competition by mouse, PDL humanized, and CDR-only humanized anti-Tac antibodies with binding of radioiodinated mouse anti-Tac antibody to HUT-102 cells.

Fig. 14. Scheme for anchored polymerase chain reaction (PCR) cloning of the heavy and light chain variable domain cDNAs. RNA was prepared from about 10^7 hybridoma cells using the hot phenol extraction method. Briefly, cells were resuspended and vortexed in 1 ml of RNA extraction buffer (50 mM sodium acetate pH 5.2/1% SDS), extracted with 0.5 ml of phenol pH 5.2 at 65°C for 15 min, followed by another 15 min on ice. The aqueous phase was recovered and precipitated twice with ethanol. cDNA was synthesized from 10 µg of total RNA using reverse transcriptase (BRL, Betheseda, MD) and oligo-dT_{12-18} (Pharmacia, Piscatway, New Jersey) as primers. A poly(dG) tail was attached to the 3' end of the cDNA using terminal deoxynucleotide transferase (BRL) (E.Y. Loh et al., Science 243, 217 (1989)), the variable domain genes (V) were amplified using AmpliTaq (Perkin Elmer-Cetus) with the primer mc045 (TAATCTAGAATTCCCCCCCCCCCCCCCCC) (SEQ ID NO:11) that hybridized to the poly(dG) tails and primers that hybridized to the constant region genes (C). For the light chain, the primer used was mc045 (TATAGAGCTCAAGCTTGGATGGTGGGAAGATGGATACAGTTGGTGC) (SEQ ID NO:12). For the heavy chain, the primer used was mc047 (TATAGAGCTCAAGCTTCCAGTGGATAGAC(CAT)GATGGGG(GC)TGT(TC)GTTTTGGC) (SEQ ID NO:13). The sequence in parenthesis indicates a base degeneracy. The degeneracy was introduced so that the primer would be able to hybridize to most gamma chains. The amplified fragments were then digested with EcoRI and HindIII and cloned into pUC18 vector for sequencing.

Fig. 15. Comparison of sequences of anti-Tac heavy chain (upper lines) (SEQ ID NO:14) and Eu heavy chain (lower lines) (SEQ ID NO:15). The 1-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions

for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

Fig. 16. Comparison of sequences of anti-Tac light chain (upper lines) (SEQ ID NO:16) and Eu light chain (lower lines) (SEQ ID NO:17). The single-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

Fig. 17. Nucleotide sequence of the gene for the humanized anti-Tac heavy chain variable region gene (SEQ ID NOS:18 and 19). The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature heavy chain sequence begins with amino acid #20 Q.

Fig. 18. Nucleotide sequence of the gene for the humanized anti-Tac light chain variable region gene (SEQ ID NOS:20 and 21). The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature light chain sequence begins with amino acid #21 D.

Fig. 19A and Fig. 19B. (A) Sequences of the four oligonucleotides (SEQ ID NOS:22, 23, 24, and 25) used to synthesize the humanized anti-Tac heavy chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide.

Fig. 20A and Fig. 20B. (A) Sequences of the four oligonucleotides (SEQ ID NOS:26, 27, 28, and 29) used to synthesize the humanized anti-Tac light chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide. The position of a Hind III site in the overlap of JFD2 and JFD3 is shown.

Fig. 21. Schematic diagram of the plasmid pHuGTAC1 used to express the humanized anti-Tac heavy chain. Relevant restriction sites are shown, and coding regions of the heavy chain are displayed as boxes. The direction of transcription from the immunoglobulin (Ig) promoter is shown by an arrow. E_H = heavy chain enhancer, Hyg = hygromycin resistance gene.

Fig. 22. Schematic diagram of the plasmid pHuLTAC used to express the humanized anti-Tac light chain. Relevant restriction sites are shown, and coding regions of the light chain are displayed as boxes. The direction

of transcription from the Ig promoter is shown by an arrow.

Fig. 23A and Fig. 23B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) (SEQ ID NOS:30 and 31) and heavy chain (B) (SEQ ID NOS:32 and 33) variable regions of the antibody mik-β1. The CDR sequences are underlined. The mature light chain protein begins with amino acid 23 Q and the mature heavy chain protein with amino acid 20 Q, preceded by the respective signal sequences.

Fig. 24A and Fig. 24B. Schematic diagram of the plasmids pVg1-dhfr (A) and pVk (B). The plasmid pVg1-dhfr contains the following parts: an approximately 4200 base pair BamHI-EcoRI fragment containing the amp and dhfr genes; a 630-bp fragment containing the human cytomegalovirus IE1 gene promoter and enhancer (Boshart et al., Cell 41, 521 (1985), which is incorporated herein by reference) flanked at the 5' and 3' ends by EcoRI and XbaI linkers respectively; and a 2800 bp XbaI-BamHI fragment containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 gene and the gpt gene replacing the dhfr gene. The plasmids were constructed from the indicated parts using methods well-known in the art (see, Maniatis et al., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1989) and commonly assigned PCT Publication No. WO 89/09622, published October 19, 1989. For example, pVg1-dhfr was constructed from the plasmid pVg1 by replacing the Hind III-Bgl II fragment containing the hyg gene with a 660 bp fragment containing the dhfr gene and extending to a Bgl II site (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983)).

Fig. 25. Fluorocytometry of YTJB cells stained with (___) Isotype matched control antibody, (---) humanized mik-β1 antibody, (...) chimeric mik-β1 antibody. Cells were suspended in FACS buffer (PBS + 2% BSA + 0.1% azide) at approximately $5x10^6$/ml. 100 µl of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with goat anti-human Ig antibody on ice for another 30 min. Then the cells were washed and incubated with FITC labeled rabbit anti-goat Ig antibody for 30 min. The cells were washed again and finally resuspended in PBS + 1% paraformaldehyde. Cells were analyzed on a FACSmate (Becton Dickinson).

Fig. 26A and Fig. 26B. Amino acid sequences of the light chain (A) (SEQ ID NOS:34 and 35) and the heavy chain (B) (SEQ ID NOS:36 and 37) of the humanized mik-β1 antibody, (lower lines) and human Lay antibody (upper lines), not including signal sequences. The three CDRs in each chain are underlined. Amino acids in the framework that have been replaced with mouse amino acids or consensus human amino acids in the humanized antibody are

double underlined.

Fig. 27A and Fig. 27B. Oligonucleotides used in the construction of the humanized mik-β1 heavy chain (B) (SEQ ID NOS:42, 43, 44, and 45) and light chain (A) (SEQ ID NOS:38, 39, 40, and 41). The following pairs of oligonucleotides were mixed, extended with sequenase and cut with the indicated enzymes before ligation into the pBluescriptII ks (+) vector: wps54 and vc11 with Xba I and Sal I, vc12 and wps57 with Xba I and Sal I, vc16 and vc13 with Xba I and Kpn I, vc14 and vc15 with Xba I and Kpn I. Then the wps54-vc11 and vc12-wps57 fragments were excised with Xba I and Sal I ligated together into the Xba I site of pVg1-dhfr; and the vc16-vc13 fragments and vc14-vc15 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVk.

Fig. 28. Competitive binding of labeled mik-β1 tracer to YTJB cells. About 10^6 YTJB cells were incubated with 3.0 ng of radio-iodinated mouse mik-β1 antibody (6 μCi/μg) and varying amounts of either unlabeled mouse mik-β1 antibody (●) or humanized mik-β1 antibody (○) in 200 μl of binding buffer (PBS + 10% fetal calf serum + 0.1% NaN_3 + 10 μg/ml mouse monoclonal Ig). After incubation for 2 hr at 0°C the cells were washed twice with binding buffer without mouse Ig and collected by centrifugation. The radioactivity bound to cells was measured and expressed as the ratio of bound/free cpm.

Fig. 29. Inhibition of IL-2 stimulated proliferation of human PHA blasts by humanized mik-β1 + humanized anti-Tac antibodies. No antibody added (□), 2 μg each of humanized mik-β1 and humanized anti-Tac added (■).

Fig. 30A through Fig. 30D. Amino acid sequences of the heavy chain (A) (SEQ ID NOS:48 and 49) and the light chain (B) (SEQ ID NOS:46 and 47) of the murine and humanized Fd79 antibodies, and the heavy chain (C) (SEQ ID NOS:52 and 53) and the light chain (D) (SEQ ID NOS:50 and 51) of the murine and humanized Fd138-80 antibodies. The sequences of the murine antibody as deduced from the cDNA (upper lines) are shown aligned with the humanized antibody sequences (lower lines). The humanized Fd79 and Fd138-80 framework sequences are derived from Pom antibody and Eu antibody, respectively. Residues are numbered according to the Kabat system (E.A. Kabat et al., Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, MD) (1987). The three CDRs in each chain are boxed. Residues in the Pom or Eu framework that have been replaced with murine sequences or consensus human sequences are underlined.

Fig. 31A and Fig. 31B. Fluorocytometry of HSV-1 infected Vero cells stained with Fd79 (A) and Fd138-80 (B) antibodies. (. .) Isotype matched control antibody, (...) humanized antibody, (___) chimeric antibody. Vero cells were infected with HSV-1 (Δ305 mutant (F strain)) at 3 pfu/cell

overnight. Cells were trypsinized at 0.5 mg/ml for 1 minute, washed extensively with PBS and resuspended in FACS buffer (PBS + 2% BSA + 0.1% azide) at approximately $5x10^6$/ml. 100 µl of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat anti-human antibody (Cappel) on ice for another 30 min. The cells were washed again and finally resuspended in PBS + 1% paraformaldehyde. Cells were analyzed on a FACSmate (Becton Dickinson).

Fig. 32A and Fig. 32B. Neutralization of HSV-1 by Fd79 (A) and Fd138-80 (B). Serial dilutions of antibodies were mixed with 100 pfu of virus and incubated at 37°C for 1 hr. The viruses were then inoculated onto 6-well plates with confluent Vero cells and adsorbed at 37°C for 1 hr. Cells were overlayed with 1% agarose in medium and incubated for 4 days. Plaques were stained with neutral red.

Fig. 33A and Fig. 33B. Immunostaining of infected Vero cell monolayers to examine protection of cells from viral spread in tissue culture by (A) murine or humanized Fd79, (B) murine or humanized Fd138-80. 24-well plates of confluent Vero cells were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37°C before adding 200 µl of 10 µg/ml antibodies in medium. At the end of 4 days, culture medium was removed and plates were dried by placing overnight in a 37°C incubator. To detect viral antigens, each well was incubated with 200 µl of anti-gB antibody at 0.5 µg/ml for 1 hr. at 37°C, washed twice and incubated with 200 µl of peroxidase conjugated goat anti-mouse IgG (Cappel, 1:300 dilution) for 1 hr. at 37°C. The plates were washed and then developed with the substrate 3-amino-9-ethyl-carbazole (AEC) (Sigma, St. Louis, MO) for 15 minutes at room temperature. Reaction was stopped by rinsing with water and air dried.

Fig. 34A and Fig. 34B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) (SEQ ID NOS:66 and 67) and heavy chain (B) (SEQ ID NOS:68 and 69) variable regions of the antibody M195. The CDR sequences are underlined. The mature light chain protein begins with amino acid 21 D and the mature heavy chain protein with amino acid 20 E, preceded by the respective signal sequences.

Fig. 35. Fluorocytometry of U937 cells stained with (. .) no antibody, (...) humanized M195 antibody, (---) chimeric M195 antibody. Cells were suspended in FACS buffer (PBS + 2% FCS + 0.1% azide) at approximately $5x10^6$/ml. 100 µl of cell suspension was transferred to a polystyrene tube and incubated with 50 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat anti-human Ig antibody on ice for another 30 min. The cells were washed again and finally resuspended in PBS + 1% paraformaldehyde. Cells

were analyzed on a FACSmate (Becton Dickinson).

Fig. 36A and Fig. 36B. Amino acid sequences of the light chain (A) (SEQ ID NOS:70 and 71) and the heavy chain (B) (SEQ ID NOS:72 and 73) of the humanized M195 antibody (lower lines) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have been replaced with mouse amino acids in the humanized antibody are double underlined.

Fig. 37A and Fig. 37B. Oligonucleotides used in the construction of the humanized M195 heavy chain (A; ma1-4) (SEQ ID NOS:74, 75, 76, and 77) and light chain (B; ma5-8) (SEQ ID NOS:78, 79, 80, and 81). The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: ma1 and ma2 with Xba I and Kpn I, ma3 and ma4 with Xba I and Kpn I, ma5 and ma6 with Xba I and Hind III, ma7 and ma8 with Xba I and Hind III. Then the ma1-ma2 and ma3-ma4 fragments were excised from pUC18 with Xba I and kpn I and ligated together into the Xba I site of pVg1-dhfr; and the ma5-ma6 and ma7-ma8 fragments were excised with Xba I and Hind III and ligated together into the Xba I site of pVk.

Fig. 38. Competitive binding of labeled M195 tracer to U937 cells. About 4 x 10^5 U937 cells were incubated with 4.5 ng of radio-iodinated mouse M195 antibody (6 μCi/μg) and varying amounts of either unlabeled mouse M195 antibody (•) or humanized M195 antibody (O) in 200 μl of binding buffer (PBS + 2% fetal calf serum + 0.1% sodium azide). After incubation for 2 hr at 0°C, the cells were washed twice with binding buffer and collected by centrifugation. The radioactivity bound to cells was measured and is expressed as the ratio of bound/free cpm.

Fig. 39A and Fig. 39B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) (SEQ ID NOS:82 and 83) and heavy chain (B) (SEQ ID NOS:84 and 85) variable regions of the antibody CMV5. The CDR sequences are underlined. The start of the mature protein sequences are indicated by arrows, preceded by the respective signal sequences.

Fig. 40A and Fig. 40B. Amino acid sequences of the light chain (A) (SEQ ID NOS:86 and 87) and the heavy chain (B) (SEQ ID NOS:88 and 89) of the humanized CMV5 antibody (lower lines) and human Wol antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

Fig. 41A and Fig. 41B. Oligonucleotides used in the construction of the humanized CMV5 light chain (A; jb16-jb19) (SEQ ID NOS:90, 91, 92, and 93) and heavy chain (B; jb20-jb22) (SEQ ID NOS:94, 95, 96, and 97). The following pairs of oligonucleotides were mixed, extended with Klenow

polymerase and cut with the indicated enzymes before ligation into pUC18: jb16 and jb17 with Xba I and EcoR I, jb18 and jb19 with Xba I and EcoR I, jb20 and jb21 with Xba I and Kpn I, jb22 and jb23 with Xba I and Kpn I. Then the jb16-jb17 and jb18-jb19 fragments were excised with Xba I and Mlu I and ligated together into the Xba I site of pVk; and the jb20-jb21 and jb22-jb23 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVgl-dhfr.

Fig. 42. Competitive binding of labeled CMV5 tracer to CMV-infected cells. Increasing amounts of mouse (•) or humanized (O) CMV5 antibody was added to CMV-infected HEL cells with tracer radio-iodinated mouse CMV5, and the amount of tracer bound to the cells was determined.

Fig. 43A and Fig. 43B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) (SEQ ID NOS:98 and 99) and heavy chain (B) (SEQ ID NOS:100 and 101) variable regions of the antibody AF2. The CDR sequences are underlined. The mature light chain protein begins with amino acid 30 N and the mature heavy chain protein with amino acid 36 Q, preceded by the respective signal sequences.

Fig. 44A and Fig. 44B. Amino acid sequences of the light chain (A) (SEQ ID NOS:102 and 103) and the heavy chain (B) (SEQ ID NOS:104 and 105) of the humanized AF2 antibody (lower lines) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have been replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

Fig. 45A and Fig. 45B. Oligonucleotides used in the construction of the humanized AF2 light chain (A; rh10-rh13) (SEQ ID NOS:106, 107, 108, and 109) and heavy chain (B; rh20-23) (SEQ ID NOS:110, 111, 112, and 113). The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: rh10 and rh11 with Xba I and Hind III, rh12 and rh13 with Xba I and Hind III, rh20 and rh21 with Xba I and EcoR I, rh22 and rh23 with Xba I and EcoR I. Then the rh10-rh11 and rh12-rh13 fragments were excised with Xba I and Hind III and ligated together into then Xba I site of pVk; and the rh20-rh21 and rh22-rh23 fragments were excised with Xba I and Xho I and ligated together into the Xba I site of pVgl-dhfr.

Fig. 46. Fluorescence of HS294T cells incubated with γ-IFN plus varying concentrations of mouse AF2 antibody, and stained with an anti-HLA-D antibody.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with the present invention, novel means of designing humanized immunoglobulins capable of specifically binding to a predetermined

antigen with strong affinity are provided. These improved methods produce immunoglobulins that are substantially non-immunogenic in humans but have binding affinities of at least about 10^8 M^{-1}, preferably 10^9 M^{-1} to 10^{10} M^{-1}, or stronger. The humanized immunoglobulins will have a human framework and have one or more complementary determining regions (CDR's), plus a limited number of other amino acids, from a donor immunoglobulin specifically reactive with an antigen. The immunoglobulins can be produced economically in large quantities and find use, for example, in the treatment of various human disorders by a variety of techniques.

In order that the invention may be more completely understood, several definitions are set forth. As used herein, the term "immunoglobulin" refers to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma (IgG_1, IgG_2, IgG_3, IgG_4), delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. Full-length immunoglobulin "light chains" (about 25 Kd or 214 amino acids) are encoded by a variable region gene at the NH2-terminus (about 110 amino acids) and a kappa or lambda constant region gene at the COOH - terminus. Full-length immunoglobulin "heavy chains" (about 50 Kd or 446 amino acids), are similarly encoded by a variable region gene (about 116 amino acids) and one of the other aforementioned constant region genes, e.g., gamma (encoding about 330 amino acids).

One form of immunoglobulin constitutes the basic structural unit of an antibody. This form is a tetramer and consists of two identical pairs of immunoglobulin chains, each pair having one light and one heavy chain. In each pair, the light and heavy chain variable regions are together responsible for binding to an antigen, and the constant regions are responsible for the antibody effector functions. In addition to antibodies, immunoglobulins may exist in a variety of other forms including, for example, Fv, Fab, and $(Fab')_2$, as well as bifunctional hybrid antibodies (e.g., Lanzavecchia et al., Eur. J. Immunol. 17, 105 (1987)) and in single chains (e.g., Huston et al., Proc. Natl. Acad. Sci. U.S.A., 85, 5879-5883 (1988) and Bird et al., Science, 242, 423-426 (1988), which are incorporated herein by reference). (See, generally, Hood et al., "Immunology", Benjamin, N.Y., 2nd ed. (1984), and Hunkapiller and Hood, Nature, 323, 15-16 (1986), which are incorporated herein by reference).

An immunoglobulin light or heavy chain variable region consists of a "framework" region interrupted by three hypervariable regions, also called CDR's. The extent of the framework region and CDR's have been precisely defined (see, "Sequences of Proteins of Immunological Interest," E. Kabat et al., U.S. Department of Health and Human Services, (1983); which is incorporated herein by reference). The sequences of the framework regions of different light or heavy chains are relatively conserved within a species. As used herein, a "human framework region" is a framework region that is substantially identical (about 85% or more, usually 90-95% or more) to the framework region of a naturally occurring human immunoglobulin. The

framework region of an antibody, that is the combined framework regions of the constituent light and heavy chains, serves to position and align the CDR's. The CDR's are primarily responsible for binding to an epitope of an antigen.

Chimeric antibodies are antibodies whose light and heavy chain genes have been constructed, typically by genetic engineering, from immunoglobulin variable and constant region genes belonging to different species. For example, the variable segments of the genes from a mouse monoclonal antibody may be joined to human constant segments, such as gamma 1 and gamma 3. A typical therapeutic chimeric antibody is thus a hybrid protein composed of the variable or antigen-binding domain from a mouse antibody and the constant or effector domain from a human antibody (e.g., A.T.C.C. Accession No. CRL 9688 secretes an anti-Tac chimeric antibody), although other mammalian species may be used.

As used herein, the term "humanized" immunoglobulin refers to an immunoglobulin comprising a human framework region and one or more CDR's from a non-human (usually a mouse or rat) immunoglobulin. The non-human immunoglobulin providing the CDR's is called the "donor" and the human immunoglobulin providing the framework is called the "acceptor". Constant regions need not be present, but if they are, they must be substantially identical to human immunoglobulin constant regions, i.e., at least about 85-90%, preferably about 95% or more identical. Hence, all parts of a humanized immunoglobulin, except possibly the CDR's, are substantially identical to corresponding parts of natural human immunoglobulin sequences. A "humanized antibody" is an antibody comprising a humanized light chain and a humanized heavy chain immunoglobulin. For example, a humanized antibody would not encompass a typical chimeric antibody as defined above, e.g., because the entire variable region of a chimeric antibody is non-human. One says that the donor antibody has been "humanized", by the process of "humanization", because the resultant humanized antibody is expected to bind to the same antigen as the donor antibody that provides the CDR's.

It is understood that the humanized antibodies designed by the present method may have additional conservative amino acid substitutions which have substantially no effect on antigen binding or other immunoglobulin functions. By conservative substitutions is intended combinations such as gly, ala; val, ile, leu; asp, glu; asn, gln; ser, thr; lys, arg; and phe, tyr.

Humanized immunoglobulins, including humanized antibodies, have been constructed by means of genetic engineering. Most humanized immunoglobulins that have been previously described (Jones et al., op. cit.; Verhoeyen et al., op. cit.; Riechmann et al., op. cit.) have comprised a framework that is identical to the framework of a particular human immunoglobulin chain, the acceptor, and three CDR's from a non-human donor immunoglobulin chain. In one case (Riechmann et al., op. cit.), two additional amino acids in the framework were changed to be the same as amino acids in other human framework regions. The present invention includes

criteria by which a limited number of amino acids in the framework of a humanized immunoglobulin chain are chosen to be the same as the amino acids at those positions in the donor rather than in the acceptor, in order to increase the affinity of an antibody comprising the humanized immunoglobulin chain.

The present invention is based in part on the model that two contributing causes of the loss of affinity in prior means of producing humanized antibodies (using as examples mouse antibodies as the source of CDR's) are:

(1) When the mouse CDR's are combined with the human framework, the amino acids in the framework close to the CDR's become human instead of mouse. Without intending to be bound by theory, we believe that these changed amino acids may slightly distort the CDR's, because they create different electrostatic or hydrophobic forces than in the donor mouse antibody, and the distorted CDR's may not make as effective contacts with the antigen as the CDR's did in the donor antibody;

(2) Also, amino acids in the original mouse antibody that are close to, but not part of, the CDR's (i.e., still part of the framework), may make contacts with the antigen that contribute to affinity. These amino acids are lost when the antibody is humanized, because all framework amino acids are made human.

To avoid these problems, and to produce humanized antibodies that have a very strong affinity for a desired antigen, the present invention uses one or more of the following principles for designing humanized immunoglobulins. Also, the criteria may be used singly, or when necessary in combination, to achieve the desired affinity or other characteristics.

A principle is that as acceptor, a framework is used from a particular human immunoglobulin that is unusually homologous to the donor immunoglobulin to be humanized, or use a consensus framework from many human antibodies. For example, comparison of the sequence of a mouse heavy (or light) chain variable region against human heavy (or light) variable regions in a data bank (for example, the National Biomedical Research Foundation Protein Identification Resource) shows that the extent of homology to different human regions varies greatly, typically from about 40% to about 60-70%. By choosing as the acceptor immunoglobulin one of the human heavy (respectively light) chain variable regions that is most homologous to the heavy (respectively light) chain variable region of the donor immunoglobulin, fewer amino acids will be changed in going from the donor immunoglobulin to the humanized immunoglobulin. Hence, and again without intending to be bound by theory, it is believed that there is a smaller chance of changing an amino acid near the CDR's that distorts their conformation. Moreover, the precise overall shape of a humanized antibody comprising the humanized immunoglobulin chain may more closely resemble the shape of the donor antibody, also reducing the chance of distorting the CDR's.

Typically, one of the 3-5 most homologous heavy chain variable region sequences in a representative collection of at least about 10 to 20

distinct human heavy chains will be chosen as acceptor to provide the heavy chain framework, and similarly for the light chain. Preferably, one of the 1-3 most homologous variable regions will be used. The selected acceptor immunoglobulin chain will most preferably have at least about 65% homology in the framework region to the donor immunoglobulin.

In many cases, it may be considered preferable to use light and heavy chains from the same human antibody as acceptor sequences, to be sure the humanized light and heavy chains will make favorable contacts with each other. In this case, the donor light and heavy chains will be compared only against chains from human antibodies whose complete sequence is known, e.g., the Eu, Lay, Pom, Wol, Sie, Gal, Ou and WEA antibodies (Kabat et al., op. cit.; occasionally, the last few amino acids of a human chain are not known and must be deduced by homology to other human antibodies). The human antibody will be chosen in which the light and heavy chain variable regions sequences, taken together, are overall most homologous to the donor light and heavy chain variable region sequences. Sometimes greater weight will be given to the heavy chain sequence. The chosen human antibody will then provide both light and heavy chain acceptor sequences. In practice, it is often found that the human Eu antibody will serve this role.

Regardless of how the acceptor immunoglobulin is chosen, higher affinity may be achieved by selecting a small number of amino acids in the framework of the humanized immunoglobulin chain to be the same as the amino acids at those positions in the donor rather than in the acceptor. A second principle is that the following categories define what amino acids may be selected from the donor. Preferably, at many or all amino acid positions in one of these categories, the donor amino acid will in fact be selected.

Category 1: The amino acid position is in a CDR is defined by Kabat et al., op. cit.

Category 2: If an amino acid in the framework of the human acceptor immunoglobulin is unusual (i.e., "rare", which as used herein indicates an amino acid occurring at that position in less than about 20% but usually less than about 10% of human heavy (respectively light) chain V region sequences in a representative data bank), and if the donor amino acid at that position is typical for human sequences (i.e., "common", which as used herein indicates an amino acid occurring in more than about 25% but usually more than about 50% of sequences in a representative data bank), then the donor amino acid rather than the acceptor may be selected. This criterion helps ensure that an atypical amino acid in the human framework does not disrupt the antibody structure. Moreover, by replacing an unusual amino acid with an amino acid from the donor antibody that happens to be typical for human antibodies, the humanized antibody may be made less immunogenic.

All human light and heavy chain variable region sequences are respectively grouped into "subgroups" of sequences that are especially homologous to each other and have the same amino acids at certain critical positions (Kabat et al., op. cit.). When deciding whether an amino acid in

a human acceptor sequence is "rare" or "common" among human sequences, it will often be preferable to consider only those human sequences in the same subgroup as the acceptor sequence.

Category 3: In the positions immediately adjacent to one or more of the 3 CDR's in the primary sequence of the humanized immunoglobulin chain, the donor amino acid(s) rather than acceptor amino acid may be selected. These amino acids are particularly likely to interact with the amino acids in the CDR's and, if chosen from the acceptor, to distort the donor CDR's and reduce affinity. Moreover, the adjacent amino acids may interact directly with the antigen (Amit et al., Science, 233, 747-753 (1986), which is incorporated herein by reference) and selecting these amino acids from the donor may be desirable to keep all the antigen contacts that provide affinity in the original antibody.

Category 4: A 3-dimensional model, typically of the original donor antibody, shows that certain amino acids outside of the CDR's are close to the CDR's and have a good probability of interacting with amino acids in the CDR's by hydrogen bonding, Van der Waals forces, hydrophobic interactions, etc. At those amino acid positions, the donor immunoglobulin amino acid rather than the acceptor immunoglobulin amino acid may be selected. Amino acids according to this criterion will generally have a side chain atom within about 3 angstrom units of some atom in the CDR's and must contain an atom that could interact with the CDR atoms according to established chemical forces, such as those listed above.

In the case of atoms that may form a hydrogen bond, the 3 angstroms is measured between their nuclei, but for atoms that do not form a bond, the 3 angstroms is measured between their Van der Waals surfaces. Hence, in the latter case, the nuclei must be within about 6 angstroms (3 + sum of the Van der Waals radii) for the atoms to be considered capable of interacting. In many cases the nuclei will be from 4 or 5 to 6A apart. In determining whether an amino acid can interact with the CDRs, it is preferred not to consider the last 8 amino acids of heavy chain CDR 2 as part of the CDRs, because from the viewpoint of structure, these 8 amino acids behave more as part of the framework.

Amino acids in the framework that are capable of interacting with amino acids in the CDR's, and which therefore belong to Category 4, may be distinguished in another way. The solvent accessible surface area of each framework amino acid is calculated in two ways: (1) in the intact antibody, and (2) in a hypothetical molecule consisting of the antibody with its CDRs removed. A significant difference between these numbers of about 10 square angstroms or more shows that access of the framework amino acid to solvent is at least partly blocked by the CDRs, and therefore that the amino acid is making contact with the CDRs. Solvent accessible surface area of an amino acid may be calculated based on a 3-dimensional model of an antibody, using algorithms known in the art (e.g., Connolly, J. Appl. Cryst. 16, 548 (1983) and Lee and Richards, J. Mol. Biol. 55, 379 (1971), both of which are

incorporated herein by reference). Framework amino acids may also occasionally interact with the CDR's indirectly, by affecting the conformation of another framework amino acid that in turn contacts the CDR's.

The amino acids at several positions in the framework are known to be capable of interacting with the CDRs in many antibodies (Chothia and Lesk, J. Mol. Biol. 196, 901 (1987), Chothia et al., Nature 342, 877 (1989), and Tramontano et al., J. Mol. Biol. 215, 175 (1990), all of which are incorporated herein by reference), notably at positions 2, 48, 64 and 71 of the light chain and 26-30, 71 and 94 of the heavy chain (numbering according to Kabat, op. cit.), and therefore these amino acids will generally be in Category 4. Typically, humanized immunoglobulins, of the present invention will include donor amino acids (where different) in category 4 in addition to these. The amino acids at positions 35 in the light chain and 93 and 103 in the heavy chain are also likely to interact with the CDRs. At all these numbered positions, choice of the donor amino acid rather than the acceptor amino acid (when they differ) to be in the humanized immunoglobulin is preferred. On the other hand, certain positions that may be in Category 4 such as the first 5 amino acids of the light chain may sometimes be chosen from the acceptor immunoglobulin without loss of affinity in the humanized immunoglobulin.

Chothia and Lesk (op. cit.) define the CDRs differently from Kabat et al. (op. cit.). Notably, CDR1 is defined as including residues 26-32. Accordingly, Riechmann et al., (op. cit.) chose these amino acids from the donor immunoglobulins.

Computer programs to create models of proteins such as antibodies are generally available and well known to those skilled in the art (see, Levy et al., Biochemistry, 28, 7168-7175 (1989); Bruccoleri et al., Nature, 335, 564-568 (1988); Chothia et al., Science, 233, 755-758 (1986), all of which are incorporated herein by reference). These do not form part of the invention. Indeed, because all antibodies have similar structures, the known antibody structures, which are available from the Brookhaven Protein Data Bank, can be used if necessary as rough models of other antibodies. Commercially available computer programs can be used to display these models on a computer monitor, to calculate the distance between atoms, and to estimate the likelihood of different amino acids interacting (see, Ferrin et al., J. Mol. Graphics, 6, 13-27 (1988)).

In addition to the above categories, which describe when an amino acid in the humanized immunoglobulin may be taken from the donor, certain amino acids in the humanized immunoglobulin may be taken from neither the donor nor acceptor, if then fall in:

Category 5: If the amino acid at a given position in the donor immunoglobulin is "rare" for human sequences, and the amino acid at that position in the acceptor immunoglobulin is also "rare" for human sequences, as defined above, then the amino acid at that position in the humanized immunoglobulin may be chosen to be some amino acid "typical" of human

sequences. A preferred choice is the amino acid that occurs most often at that position in the known human sequences belonging to the same subgroup as the acceptor sequence.

Humanized antibodies generally have at least three potential advantages over mouse or in some cases chimeric antibodies for use in human therapy:

1) Because the effector portion is human, it may interact better with the other parts of the human immune system (e.g., destroy the target cells more efficiently by complement-dependent cytotoxicity (CDC) or antibody-dependent cellular cytotoxicity (ADCC)).

2) The human immune system should not recognize the framework or constant region of the humanized antibody as foreign, and therefore the antibody response against such an injected antibody should be less than against a totally foreign mouse antibody or a partially foreign chimeric antibody.

3) Injected mouse antibodies have been reported to have a half-life in the human circulation much shorter than the half-life of normal antibodies (D. Shaw et al., J. Immunol., 138, 4534-4538 (1987)). Injected humanized antibodies will presumably have a half-life more similar to naturally occurring human antibodies, allowing smaller and less frequent doses to be given.

In one aspect, the present invention is directed to designing humanized immunoglobulins that are produced by expressing recombinant DNA segments encoding the heavy and light chain CDR's from a donor immunoglobulin capable of binding to a desired antigen, such as the human IL-2 receptor, attached to DNA segments encoding acceptor human framework regions. Exemplary DNA sequences designed in accordance with the present invention code for the polypeptide chains comprising heavy and light chain CDR's with substantially human framework regions shown in Fig. 1A through Fig. 6B. Due to codon degeneracy and non-critical amino acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. In general, the criteria of the present invention find applicability to designing substantially any humanized immunoglobulin.

The DNA segments will typically further include an expression control DNA sequence operably linked to the humanized immunoglobulin coding sequences, including naturally-associated or heterologous promoter regions. Preferably, the expression control sequences will be eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells, but control sequences for prokaryotic hosts may also be used. Once the vector has been incorporated into the appropriate host, the host is maintained under conditions suitable for high level expression of the nucleotide sequences, and, as desired, the collection and purification of the humanized light chains, heavy chains, light/heavy chain dimers or intact antibodies, binding fragments or other immunoglobulin forms may follow (see, S. Beychok, Cells of Immunoglobulin Synthesis, Academic Press, N.Y., (1979), which is incorporated herein by reference).

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to the predetermined antigen, such as the human IL-2 receptor, and produced by well known methods in any convenient mammalian source including, mice, rats, rabbits, or other vertebrates, capable of producing antibodies. Suitable source cells for the constant region and framework DNA sequences, and host cells for immunoglobulin expression and secretion, can be obtained from a number of sources, such as the American Type Culture Collection ("Catalogue of Cell Lines and Hybridomas," sixth edition (1988) Rockville, Maryland, U.S.A., which is incorporated herein by reference).

In addition to the humanized immunoglobulins specifically described herein, other "substantially homologous" modified immunoglobulins to the native sequences can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the framework regions can vary specifically from the sequences in Fig. 1A through Fig. 6B at the primary structure level by several amino acid substitutions, terminal and intermediate additions and deletions, and the like. Moreover, a variety of different human framework regions may be used singly or in combination as a basis for the humanized immunoglobulins of the present invention. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gillman and Smith, Gene, 8, 81-97 (1979) and S. Roberts et al., Nature, 328, 731-734 (1987), both of which are incorporated herein by reference).

Substantially homologous immunoglobulin sequences are those which exhibit at least about 85% homology, usually at least about 90%, and preferably at least about 95% homology with a reference immunoglobulin protein.

Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVg1 (Figs. 9A and 9B) using site-directed mutagenesis, such as after CH1 to produce Fab fragments or after the hinge region to produce $(Fab')_2$ fragments. Single chain antibodies may be produced by joining V_L and V_H with a DNA linker (see, Huston et al., op. cit., and Bird et al., op. cit.). Also because like many genes, the immunoglobulin-related genes contain separate functional regions, each having one or more distinct biological activities, the genes may be fused to functional regions from other genes (e.g., enzymes, see, commonly assigned U.S. Patent No. 5,004,692) to produce fusion proteins (e.g., immunotoxins) having novel properties. The nucleic acid sequences of the present invention capable of ultimately expressing the desired humanized

antibodies can be formed from a variety of different polynucleotides (genomic or cDNA, RNA, synthetic oligonucleotides, etc.) and components (e.g., V, J, D, and C regions), as well as by a variety of different techniques. Joining appropriate synthetic and genomic sequences is presently the most common method of production, but cDNA sequences may also be utilized (see, European Patent Publication No. 0239400 and L. Reichmann et al., Nature, 332, 323-327 (1988), both of which are incorporated herein by reference).

As stated previously, the DNA sequences will be expressed in hosts after the sequences have been operably linked to (i.e., positioned to ensure the functioning of) an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors will contain selection markers, e.g., tetracycline or neomycin, to permit detection of those cells transformed with the desired DNA sequences (see, e.g., U.S. Patent 4,704,362, which is incorporated herein by reference).

E. coli is one prokaryotic host useful particularly for cloning the DNA sequences of the present invention. Other microbial hosts suitable for use include bacilli, such as Bacillus subtilus, and other enterobacteriaceae, such as Salmonella, Serratia, and various Pseudomonas species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well- known promoters will be present, such as the lactose promoter system, a tryptophan (trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

Other microbes, such as yeast, may also be used for expression. Saccharomyces is a preferred host, with suitable vectors having expression control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired.

In addition to microorganisms, mammalian tissue cell culture may also be used to express and produce the polypeptides of the present invention (see, Winnacker, "From Genes to Clones," VCH Publishers, N.Y., N.Y. (1987), which is incorporated herein by reference). Eukaryotic cells are actually preferred, because a number of suitable host cell lines capable of secreting intact immunoglobulins have been developed in the art, and include the CHO cell lines, various COS cell lines, HeLa cells, preferably myeloma cell lines, etc, and transformed B-cells or hybridomas. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, an enhancer (Queen et al., Immunol. Rev., 89, 49-68 (1986), which is incorporated herein by reference), and necessary processing information sites, such as ribosome binding sites, RNA splice

sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, Adenovirus, cytomegalovirus, Bovine Papilloma Virus, and the like.

The vectors containing the DNA segments of interest (e.g., the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts. (See, generally, Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, (1982), which is incorporated herein by reference.)

Once expressed, the whole antibodies, their dimers, individual light and heavy chains, or other immunoglobulin forms of the present invention, can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, R. Scopes, "Protein Purification", Springer-Verlag, N.Y. (1982)). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses. Once purified, partially or to homogeneity as desired, the polypeptides may then be used therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings, and the like. (See, generally, Immunological Methods, Vols. I and II, Lefkovits and Pernis, eds., Academic Press, New York, N.Y. (1979 and 1981)).

The antibodies of the present invention will typically find use individually in treating substantially any disease susceptible to monoclonal antibody-based therapy. In particular, the immunoglobulins can be used for passive immunization or the removal of unwanted cells or antigens, such as by complement mediated lysis, all without substantial immune reactions (e.g., anaphylactic shock) associated with many prior antibodies. For example, where the cell linked to a disease has been identified as IL-2 receptor bearing, then humanized antibodies that bind to the human IL-2 receptor are suitable (see, U.S.S.N. 085,707, entitled "Treating Human Malignancies and Disorders," which is incorporated herein by reference). For such a humanized immunoglobulin, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The method of producing humanized antibodies of the present invention can be used to humanize a variety of donor antibodies, especially monoclonal antibodies reactive with markers on cells responsible for a disease. For example, suitable antibodies bind to antigens on T-cells, such as those grouped into the so-called "Clusters of Differentiation," as named

by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference.

The antibodies of the present invention can also be used as separately administered compositions given in conjunction with chemotherapeutic or immunosuppressive agents. Possible agents include cyclosporin A or a purine analog (e.g., methotrexate, 6-mercaptopurine, or the like), but numerous additional agents (e.g., cyclophosphamide, prednisone, etc.) well-known to those skilled in the art of medicine may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject antibodies in immunotoxins. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells comprising a carcinoma. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Monoclonal Antibodies in Clinical Medicine, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see, Chaudhary et al., Nature 339, 394 (1989), which is herein incorporated by reference).

A variety of cytotoxic agents are suitable for use in immunotoxins. Cytotoxic agents can include radionuclides, such as Iodine-131 or other isotopes of iodine, Yttrium-90, Rhenium-188, and Bismuth-212 or other alpha emitters; a number of chemotherapeutic drugs, such as vindesine, methotrexate, adriamycin, and cisplatinum; and cytotoxic proteins such as ribosomal inhibiting proteins like pokeweed antiviral protein, Pseudomonas exotoxin A, ricin, diphtheria toxin, ricin A chain, etc., or an agent active at the cell surface, such as the phospholipase enzymes (e.g., phospholipase C). (See, generally, "Chimeric Toxins," Olsnes and Phil, Pharmac. Ther., 25, 355-381 (1982), and "Monoclonal Antibodies for Cancer Detection and Therapy," eds. Baldwin and Byers, pp. 159-179, 224-266, Academic Press (1985), all of which are incorporated herein by reference.)

The delivery component of the immunotoxin will include the humanized immunoglobulins of the present invention. Intact immunoglobulins or their binding fragments, such as Fab, are preferably used. Typically, the antibodies in the immunotoxins will be of the human IgM or IgG isotype, but other mammalian constant regions may be utilized as desired.

For diagnostic purposes, the antibodies may either be labeled or unlabeled. Unlabeled antibodies can be used in combination with other

labeled antibodies (second antibodies) that are reactive with the humanized antibody, such as antibodies specific for human immunoglobulin constant regions. Alternatively, the antibodies can be directly labeled. A wide variety of labels may be employed, such as radionuclides, fluors, enzymes, enzyme substrates, enzyme co-factors, enzyme inhibitors, ligands (particularly haptens), etc. Numerous types of immunoassays are available and are well known to those skilled in the art.

Anti-IL-2 Receptor Antibodies

To exert its biological effects, IL-2 interacts with a specific high-affinity membrane receptor (Greene, W., et al., Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem. 58, 875 (1989), both of which are incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide, being about 55kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). A gene encoding this protein has been isolated, and predicts a 272 amino acid peptide, including a 21 amino acid signal peptide (see, Leonard, W., et al., Nature 311, 626 (1984)). The 219 NH_2-terminal amino acids of the p55 Tac protein apparently comprise an extracellular domain (see, Leonard, W., et al., Science, 230, 633-639 (1985), which is incorporated herein by reference).

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. In particular, one mouse monoclonal antibody, known as anti-Tac (Uchiyama, et al., J. Immunol. 126, 1393 (1981)) has been used to show that IL-2 receptors can be detected on T-cells, but also on cells of the monocyte-macrophage family, Kupffer cells of the liver, Langerhans' cells of the skin and, of course, activated T-cells. Importantly, resting T-cells, B-cells or circulating macrophages typically do not display the IL-2 receptor (Herrmann, et al., J. Exp. Med. 162, 1111 (1985)).

The anti-Tac monoclonal antibody has also been used to define lymphocyte functions that require IL-2 interaction, and has been shown to inhibit various T-cell functions, including the generation of cytotoxic and suppressor T lymphocytes in cell culture. Also, based on studies with anti-Tac and other antibodies, a variety of disorders are now associated with improper IL-2 receptor expression by T-cells, in particular adult T-cell leukemia.

More recently, the IL-2 receptor has been shown to be an ideal target for novel therapeutic approaches to T-cell mediated diseases. It has been proposed that IL-2 receptor specific antibodies, such as the anti-Tac monoclonal antibody, can be used either alone or as an immunoconjugate (e.g., with Ricin A, isotopes and the like) to effectively remove cells bearing the IL-2 receptor. These agents can, for example, theoretically eliminate IL-2 receptor-expressing leukemic cells, certain B-cells, or activated T-cells involved in a disease state, yet allow the retention of mature normal T-cells and their precursors to ensure the capability of

mounting a normal T-cell immune response as needed. In general, most other T-cell specific agents can destroy essentially all peripheral T-cells, which limits the agents' therapeutic efficacy. Overall, the use of appropriate monoclonal antibodies specific for the IL-2 receptor may have therapeutic utility in autoimmune diseases, organ transplantation and any unwanted response by activated T-cells. Indeed, clinical trials have been initiated using, e.g., anti-Tac antibodies (see, generally, Waldmann, T., et al., Cancer Res. 45, 625 (1985), Waldmann, T., Science 232, 727-732 (1986) and Kirkman et al., Transplant. Proc. 21, 1766 (1989), all of which are incorporated herein by reference).

Unfortunately, the use of the anti-Tac and other non-human monoclonal antibodies have certain drawbacks, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

Perhaps more importantly, anti-Tac and other non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that, after injection of a foreign antibody, the immune response elicited by a patient against an antibody can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after the first or several treatments with any different non-human antibodies, subsequent treatments even for unrelated therapies can be ineffective or even dangerous in themselves, because of cross-reactivity.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with the human IL-2 receptor, as with many human antigens, has been extremely difficult using typical human monoclonal antibody production techniques. Similarly, utilizing recombinant DNA technology to produce so-called "reshaped" or "humanized" antibodies (see, e.g., Riechmann et al., Nature 332, 323 (1988) and EPO Publication No. 0239400), provides uncertain results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of human-like immunoglobulins specific for the human IL-2 receptor that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of blocking the binding of human IL-2 to its receptor and/or capable of binding to the p55 Tac protein on human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one

chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the human IL-2 receptor at affinity levels stronger than about 10^8 M^{-1}.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for human-like immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the IL-2 receptor on human T-cells are provided. These immunoglobulins, which have binding affinities of at least about 10^8 M^{-1}, and preferably 10^9 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitope on p55 Tac protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

The basic antibody structural unit is known to comprise a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25kD) and one "heavy" chain (about 50-70kD). The NH_2-terminus of each chain begins a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. The COOH terminus of each chain defines a constant region primarily responsible for effector function.

Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta, or epsilon, and define the antibody's isotype as IgG, IgM, IgA, IgD and IgE, respectively. Within light and heavy chains, the variable and constant regions are joined by a "J" region of about 10 or more amino acids, with the heavy chain also including a "D" region of about 12 more amino acids. (See, generally,

Fundamental Immunology, Paul, W., Ed., Chapter 7, pgs. 131-166, Raven Press, N.Y. (1984), which is incorporated herein by reference.)

The variable regions of each light/heavy chain pair form the antibody binding site. The chains all exhibit the same general structure of relatively conserved framework regions joined by three hypervariable regions, also called Complementarity Determining Regions or CDR's (see, "Sequences of Proteins of Immunological Interest," Kabat, E., et al., U.S. Department of Health and Human Services, (1983); and Chothia and Lesk, J. Mol. Biol., 196, 901-917 (1987), which are incorporated herein by reference). The CDR's from the two chains of each pair are aligned by the framework regions, enabling binding to a specific epitope.

As used herein, the term "immunoglobulin" refers to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. The immunoglobulins may exist in a variety of forms besides antibodies; including, for example, Fv, Fab, and $(Fab')_2$, as well as bifunctional hybrid antibodies (e.g., Lanzavecchia et al., Eur. J. Immunol. 17, 105 (1987)) and in single chains (e.g., Huston, et al., Proc. Nat. Acad. Sci. U.S.A., 85, 5879-5883 (1988) and Bird, et al., Science, 242, 423-426 (1988), which are incorporated herein by reference). (See, generally, Hood, et al., "Immunology", Benjamin, N.Y., 2nd ed. (1984), and Hunkapiller and Hood, Nature, 323, 15-16 (1986), which are incorporated herein by reference).

Chimeric antibodies are antibodies whose light and heavy chain genes have been constructed, typically by genetic engineering, from immunoglobulin gene segments belonging to different species. For example, the variable (V) segments of the genes from a mouse monoclonal antibody may be joined to human constant (C) segments, such as γ_1 and γ_3. A typical therapeutic chimeric antibody is thus a hybrid protein consisting of the V or antigen-binding domain from a mouse antibody and the C or effector domain from a human antibody (e.g., A.T.C.C. Accession No. CRL 9688 secretes an anti-Tac chimeric antibody), although other mammalian species may be used.

As used herein, the term "framework region" refers to those portions of immunoglobulin light and heavy chain variable regions that are relatively conserved (i.e., other than the CDR's) among different immunoglobulins in a single species, as defined by Kabat, et al., op. cit. As used herein, a "human-like framework region" is a framework region that in each existing chain comprises at least about 70-75 or more amino acid residues, typically 75 to 85 or more residues, identical to those in a human immunoglobulin.

As used herein, the term "humanized" immunoglobulin refers to an immunoglobulin comprising (1) a human-like framework, (2) at least one CDR from a non-human antibody, and (3) in which any constant region is substantially homologous to a human immunoglobulin constant region, i.e., at least about 85-90% identical, preferably at least 95% identical.

In one aspect, the present invention is directed to recombinant

DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the anti-Tac monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Preferred DNA sequences, which on expression code for the polypeptide chains comprising the anti-Tac heavy and light chain hypervariable regions (with human-like framework regions), are included in Figs. 15 and 16, respectively. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference.

The human-like antibodies and pharmaceutical compositions thereof of this invention are particularly useful for parenteral administration, i.e., subcutaneously, intramuscularly or intravenously. The compositions for parenteral administration will commonly comprise a solution of the antibody or a cocktail thereof dissolved in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers can be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine and the like. These solutions are sterile and generally free of particulate matter. These compositions may be sterilized by conventional, well known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, toxicity adjusting agents and the like, for example sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium lactate, human albumin, etc. The concentration of antibody in these formulations can vary widely, i.e., from less than about 0.5%, usually at or at least about 1% to as much as 15 or 20% by weight and will be selected primarily based on fluid volumes, viscosities, etc., in accordance with the particular mode of administration selected.

Thus, a typical pharmaceutical composition for injection could be made up to contain 1 ml sterile buffered water, and 1 to 50 mg of antibody.

A typical composition for intravenous infusion could be made up to contain 250 ml of sterile Ringer's solution, and 150 mg of antibody. Actual methods for preparing parenterally administrable compositions will be known or apparent to those skilled in the art and are described in more detail in, for example, Remington's Pharmaceutical Science, 15th ed., Mack Publishing Company, Easton, Pennsylvania (1980), which is incorporated herein by reference.

The antibodies of this invention can be frozen or lyophilized for storage and reconstituted in a suitable carrier prior to use. This technique has been shown to be effective with conventional immune globulins and art-known lyophilization and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilization and reconstitution can lead to varying degrees of antibody activity loss (e.g., with conventional immune globulins, IgM antibodies tend to have greater activity loss than IgG antibodies) and that use levels may have to be adjusted to compensate.

The compositions containing the present human-like antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from a disease, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially life-threatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present human-like antibodies of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present antibodies or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose, especially 0.5 to 2.5 mg per dose. A preferred prophylactic use is for the prevention of kidney transplant rejection.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ies) of this invention sufficient to effectively treat the patient.

Human-like antibodies of the present invention can further find a

wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Kits can also be supplied for use with the subject antibodies in the protection against or detection of a cellular activity or for the presence of a selected antigen. Thus, the subject antibody composition of the present invention may be provided, usually in a lyophilized form in a container, either alone or in conjunction with additional antibodies specific for the desired cell type. The antibodies, which may be conjugated to a label or toxin, or unconjugated, are included in the kits with buffers, such as Tris, phosphate, carbonate, etc., stabilizers, biocides, inert proteins, e.g., serum albumin, or the like, and a set of instructions for use. Generally, these materials will be present in less than about 5% wt. based on the amount of active antibody, and usually present in total amount of at least about 0.001% wt. based again on the antibody concentration. Frequently, it will be desirable to include an inert extender or excipient to dilute the active ingredients, where the excipient may be present in from about 1 to 99% wt. of the total composition. Where a second antibody capable of binding to the chimeric antibody is employed in an assay, this will usually be present in a separate vial. The second antibody is typically conjugated to a label and formulated in an analogous manner with the antibody formulations described above.

p75 Chain of IL-2 Receptor

The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide or alpha chain, being about 55kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., Proc. Nat. Acad. Sci. USA, 83, 9694 (1986) and Sharon et al., Science 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

The present invention provides novel compositions useful, for example, in the treatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of inhibiting the binding of human IL-2 to its receptor and/or capable of binding to the p75 protein of human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the p75 protein at affinity levels stronger than about 10^7 M^{-1}. These humanized immunoglobulins will also be capable of

blocking the binding of the CDR-donating mouse monoclonal antibody to p75.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the p75 chain of the human IL-2 receptor are provided. These immunoglobulins, which have binding affinities of at least 10^7 to 10^8 M^{-1}, and preferably 10^9 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitope on p75 protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the mik-β1 monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the mik-β1 heavy and light chain CDRs, are included in Fig. 23A and Fig. 23B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft-versus-host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference. A

preferred use is the simultaneous treatment of a patient with a human-like antibody binding to p55 and a human-like antibody binding to p75 of the IL-2 receptor, i.e., humanized anti-Tac plus humanized mik-β1.

Human-like antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Anti- HSV Antibodies

Herpes Simplex Virus types I and II (HSV-1 and HSV-2), are now estimated to be the second most frequent cause of sexually transmitted diseases in the world. Although completely accurate data are not available, infection estimates range from about 20 to 40% of the U.S. population.

A large number of diseases, from asymptomatic to life-threatening, are associated with HSV infection. Of particular clinical interest, encephalitis from HSV-1 infection and transmission of HSV-2 from a pregnant mother to her fetus are often fatal. Immunosuppressed patients are also subject to severe complications when infected with the virus. More than 50 HSV polypeptides have been identified in HSV-infected cells, including at least seven major cell surface glycoproteins (see, Whitley, R., Chapt. 66, and Roizman and Sears, Chapt. 65, Virology, Eds. Fields et al., 2nd ed., Raven Press, N.Y., N.Y. (1990), which are incorporated herein by reference). The specific biologic functions of these glycoproteins are not well defined, although gB and gD have been shown to be associated with cell fusion activity (W. Cai et al., J. Virol. 62, 2596 (1988) and Fuller and Spear, Proc. Natl. Acad. Sci. USA 84, 5454 (1987)). gB and gD express both type-specific and type-common antigenic determinants. Oakes and Lausch demonstrated that monoclonal antibodies against gB and gE suppress replication of HSV-1 in trigeminal ganglia (Oakes and Lausch, J. Virol. 51, 656 (1984)). Dix et al. showed that anti-gC and gD antibodies protect mice against acute virus-induced neurological disease (Dix et al., Infect. Immun. 34, 192 (1981)). Whitley and colleagues produced a panel of murine monoclonal antibodies against HSV-1 and showed that several of the antibodies protected mice against encephalitis and death following ocular inoculation with the virus (see, Koga et al., Virology 151, 385 (1986); Metcalf et al., Cur. Eye Res. 6, 173 (1987) and Metcalf et al., Intervirology 29, 39 1988), all of which are incorporated herein by reference). Clone Fd79 (anti-gB) prevented encephalitis even when immunization was delayed until 48 hours post-infection. Fd79 and Fd138-80 (anti-gD) significantly reduced the severity of epithelial keratitis and lowered the frequency of persistent viral infection in an outbred mouse model.

Thus, there is a need for improved forms of humanized immunoglobulins specific for HSV antigens that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of HSV mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of HSV to its receptors and/or capable of binding to the HSV specific proteins. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the HSV surface proteins at affinity levels stronger than about 10^7 M^{-1}. These humanized immunoglobulins will also be capable of blocking the binding of the CDR donating mouse monoclonal antibody to HSV.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with an antiviral agent, such as acyclovir or a cytotoxic agent active at viral surfaces. All of these compounds will be particularly useful in treating HSV mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with HSV related epitopes either directly on the virus or on infected cells are provided. These immunoglobulins, which have binding affinities to HSV specific antigens of at least about 10^7 M^{-1}, and preferably 10^8 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., protecting cells from HSV transmission. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an HSV protein, such as gB and gD proteins. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of HSV mediated disorders in human patients by a variety of techniques.

The HSVs are among the most intensively investigated of all viruses, and the HSV virion structure has been shown to contain about 33 proteins. Humanized immunoglobulins utilizing CDR's from monoclonal antibodies reactive with these proteins, particularly the eight surface glycoproteins (e.g., gB, gC, gD, gE, gG, gH and gI), represent preferred embodiments of the present invention (see, Spear, P.G., The Herpesviruses, vol. 3, pp. 315-356 (1984) (Roizman, B., ed), Plenum Press, N.Y., N.Y. and Spear, P.G., Immunochemistry of Viruses. The Basis for Serodiagnosis and Vaccines, pp. 425-446 (1985) (Neurath, A.R., eds.), Amsterdam: Elsevier, both of which are incorporated herein by reference).

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of an HSV protein,

such as monoclonal antibodies reactive with HSV gB and gD glycoproteins. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate humanized framework regions. Exemplary DNA sequences code for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd79 and Fd138-80, shown in Fig. 30A through Fig. 30D. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating an HSV mediated disease state. For example, typical disease states suitable for treatment include any involving HSV infection. Specific diseases include neonatal herpes, herpes encephalitis, ocular herpes, genital herpes and disseminated herpes (see, Corey, L., Chapter 136, Harrison's Principles of Internal Medicine, 11th ed., McGraw-Hill Book Company, N.Y., N.Y. (1987), which is incorporated herein by reference).

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different HSV antigens. For example, suitable HSV antigens to which a cocktail of humanized immunoglobulins may react include gC, gE, gF, gG and gH (see, Rector, J. et al., Infect. Immun. 38, 168 (1982) and Fuller, A. et al., J. Virol. 63, 3435 (1989), both of which are incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with acyclovir or other antiviral agents. Typically, the agents may include idoxuridine or trifluorothymidine, but numerous additional agents (e.g., vidarabine) well-known to those skilled in the art for HSV treatment may also be utilized (see, Corey, L., op. cit.).

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill cells infected by HSV. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing an HSV epitope.

The compositions containing the present humanized antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from HSV infection, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially life-

threatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present humanized immunoglobulins of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose. A preferred prophylactic use is for the prevention of herpes in immunocompromised patients, such as organ transplant recipients.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ies) of this invention sufficient to effectively treat the patient.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of HSV antigens, for isolating specific HSV infected cells or fragments of the virus, for vaccine preparation, or the like.

Anti-CD33 Antibodies

There are about 10,000-15,000 new cases of myeloid (also called non-lymphocytic or granulocytic) leukemia in the U.S. per year (Cancer Facts & Figures, American Cancer Society, 1987). There are two major forms of myeloid leukemia: acute myelogenous leukemia (AML) and chronic myelogenous leukemia (CML). Despite treatment with chemotherapy, long-term survival in patients with AML is less than 10-20% (Clarkson et al., CRC Critical Review in Oncology/Hematology 4, 221 (1986)), and survival with CML and related diseases such as chronic myelomonocytic leukemia (CMML), chronic monocytic leukemia (CMMOL) and myelodysplastic syndrome (MDS) is even lower.

The p67 protein or CD33 antigen is found on the surface of progenitors of myeloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or non-hematopoietic cells (see, Leucocyte Typing III, ed. by A.J. McMichael, Oxford University Press, pp. 622-629 (1987), which is incorporated herein by reference). Antibodies that are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

Another antibody that binds to CD33 is M195 (Tanimoto et al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference). The reactivity of M195 with a wide variety of cells and tissues was tested. Among normal cells, M195 was reported to bind only to some monocytes and myeloid progenitor cells.

The research also reported that it does not bind to other hematopoietic cells or to non-hematopoietic tissues. M195 bound to cells of most cases of AML and all cases of CML in myeloblastic phase.

A phase I clinical trial of M195 in AML has been conducted (Scheinberg et al., Proc. ASCO 9, 207 (1990)). M195 radiolabeled with iodine-131 was found to rapidly and specifically target leukemic cells in both the blood and bone marrow.

Unfortunately, the use of non-human monoclonal antibodies such as M195 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CD33 antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of myeloid leukemia-related human disorders, the compositions containing humanized immunoglobulins specifically capable of binding to CD33 antigen. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the CD33 antigen at affinity levels stronger than about 10^7 M^{-1}. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to CD33.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such as cytosine arabinoside or daunorubicin active against leukemia cells, or complexed with a radionuclide such as iodine-131. All of these compounds will be particularly useful in treating leukemia and myeloid cell-mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CD33 related epitopes are provided. These immunoglobulins, which have binding affinities to CD33 of at least about 10^7 M^{-1}, and preferably 10^8 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., destroying leukemia cells. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with CD33 antigen. In a preferred embodiment, one or more of the CDR's will come from the M195 antibody. Importantly, M195 does not bind to the ultimate hematopoietic stem cells, so M195 used in therapy will minimally interact with and destroy those cells, which are critical for generating all blood cells. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of myeloid cell-mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of CD33 antigen, such as monoclonal antibodies M195, L4B3, L1B2 or MY9. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody M195 are included in Fig. 34A and Fig. 34B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating hematologic malignancies. For example, typical disease states suitable for treatment include AML, CML, CMML, CMMOL and MDS (see, generally, Hoffbrand & Pettit, Essential Haematology, Blackwell Scientific Publications, Oxford (1980)). The antibodies may also be used for bone marrow ablation prior to bone marrow transplant.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different myeloid antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include CD13, CD14, CD15, CD16 and CD34 (see, Leukocyte Typing III, op. cit., pp. 576-732).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include cytosine arabinoside and daunorubicin, but numerous additional agents (e.g., 6-thioguanine) well-known to those skilled in the art for leukemia treatment may also be utilized (see, Hoffbrund & Pettit., op. cit.).

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill leukemia cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when

attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a CD33 epitope.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of CD33 antigens, for isolating specific myeloid cells, or the like.

It will be understood that although examples pertain to the M195 antibody, producing humanized antibodies with high binding affinity for the CD33 antigen is also contemplated using CDR's from L4B3, L1B2, MY9 or other monoclonal antibodies that bind to an epitope of CD33.

Anti-CMV Antibodies

Cytomegalovirus is a major pathogen of immunocompromised individuals, especially bone marrow transplant recipients, organ transplant recipients, and AIDS patients (see, generally, Fields et al., Eds., Virology, 2nd ed., Raven Press, New York pp. 1981-2010 (1990), which is incorporated herein by reference). Approximately 15% of bone marrow transplant patients develop CMV pneumonia, with an 85% mortality rate (Meyers, Rev. Inf. Dis. 11 (suppl. 7), S1691 (1989)). About 10% of AIDS patients develop severe CMV disease; and congenitally acquired CMV, often with significant morbidity and mortality, affects 1% of newborns (Fields, op. cit.).

The drug ganciclovir is effective against certain forms of CMV infection, notably chorioretinitis and gastroenteritis, but is not very effective against CMV pneumonia, and it has serious toxicity. Use of pooled human immunoglobulin preparations has shown some beneficial effect for prophylaxis of CMV in bone marrow transplant patients (Meyers, op. cit.), and a combination of high-dose immune globulin and ganciclovir has been reported effective against CMV pneumonia (Emanuel et al., Trans. Proc. XIX (suppl. 7), 132 (1987)). However, the marginal effectiveness, variable potency and high cost of commercial human immune globulin remain serious problems. Hence, there is a great need for new drugs effective against CMV.

CMV is a member of the herpesvirus family of viruses, and as such, has a large double-stranded DNA core, a protein capsid, and an outer lipid envelope with viral glycoproteins on its surface. At least 8 proteins have been detected on the envelope of CMV (Britt et al., J. Virol. 62, 3309 (1988)) and others have been predicted to exist based on the DNA sequence of CMV (Chee et al., Nature 344, 774 (1990)). Murine monoclonal antibodies have been produced against two especially significant CMV glycoproteins: gB, also called p130/55 or gp55-116, and gH, also called p86 (Rasmussen et al., Virology 163, 308 (1988) and Britt et al., op. cit., both of which are incorporated herein by reference) and shown to neutralize infectivity of the virus. Three other neutralizing antibodies to gH are designated CMV5, CMV109 and CMV115. Human monoclonal antibodies to CMV have also been produced (Ehrlich et al., Hybridoma 6, 151 (1987)).

In animal models, murine monoclonal antibodies have been shown

effective in treating infections caused by various viruses, including members of the herpesvirus family (see, e.g., Metcalf et al., Intervirol. 29, 39 (1988)). Hence, such antibodies may be useful in treatment of CMV infections.

Unfortunately, the use of non-human monoclonal antibodies such as CMV5 and CMV115 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with CMV antigens, as with many antigens, is difficult using typical human monoclonal antibody production techniques. Moreover, the human antibodies produced may lack certain desirable properties, such as high binding affinity and the ability to neutralize all clinical CMV strains. Similarly, utilizing recombinant DNA technology to produce so-called "humanized" or "reshaped" antibodies (see, e.g., Riechmann et al., Nature 332, 323 (1988) and EPO Publication No. 0239400, which are incorporated herein by reference), provides uncertain results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CMV antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of CMV-mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of CMV to its receptors and/or capable of binding to CMV antigens. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to CMV at affinity levels stronger than about 10^7 M^{-1}. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to CMV.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA

segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such a acyclovir or ganciclovir active against CMV-infected cells, or complexed with a cytotoxic agent. All of these compounds will be particularly useful in treating CMV-mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CMV and CMV-infected cells are provided. These immunoglobulins, which have binding affinities to CMV specific antigens of at least about 10^7 M^{-1}, and preferably 10^8 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., blocking CMV infection of cells. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with a CMV antigen. In a preferred embodiment, one or more of the CDR's will come from the CMV5, or CMV109 or CMV115 antibodies. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of CMV-mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of a CMV antigen, such as monoclonal antibodies CMV5 or CMV115. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody CMV5 are included in Fig. 39A and Fig. 39B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit. and WO87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to CMV and produced in any convenient mammalian source, including, mice, rats, rabbits, or other vertebrate capable of producing antibodies by well known methods. Suitable source cells for the DNA sequences and host cells for immunoglobulin expression and secretion can be obtained from a number of sources, such as the American Type Culture Collection (Catalogue of Cell Lines and Hybridomas, Fifth edition (1985) Rockville, Maryland, U.S.A., which is incorporated herein by reference).

The antibodies of the present invention will typically find use individually in treating CMV-related disorders. For example, typical disease states suitable for treatment include CMV pneumonia, neonatal CMV infection, CMV mononucleosis and CMV-related chorioretinitis and

gastroenteritis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different CMV antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include the gB and gH proteins.

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include acyclovir or ganciclovir, but numerous additional agents well-known to those skilled in the art for CMV treatment may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill CMV-infected cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells *in vitro* or *in vivo*. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a CMV epitope. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, *e.g.*, SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., *Monoclonal Antibodies in Clinical Medicine*, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see Chaudhary et al., Nature *339*, 394 (1989)).

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 1 to 50 mg per dose. A preferred prophylactic use is for the prevention of CMV infection in immunocompromised patients, such as organ or bone marrow transplant recipients.

Humanized antibodies of the present invention can further find a wide variety of utilities *in vitro*. By way of example, the antibodies can be utilized for detection of CMV antigens, for isolating specific CMV-infected cells, or the like.

In particular, the same method may be used to produce a humanized CMV109, CMV115 or other anti-CMV antibody as used to produce humanized CMV5 herein.

Anti-γ-IFN Antibodies

In mammals, the immune response is mediated by several types of cells that interact specifically with foreign material, i.e., antigens. One of these cell types, B cells, is responsible for the production of antibodies. Another cell type, T cells, include a wide variety of cellular subsets that destroy virally infected cells or control the in vivo function of both B cells and other hematopoietic cells, including T cells. A third cell type, macrophages, process and present antigens in conjunction with major histocompatibility complex (MHC) proteins to T cells. Communication between these cell types is mediated in a complex manner by lymphokines, such as interleukins 1-6 and γ-IFN (see, generally, Paul, W.E., ed., Fundamental Immunology, 2nd ed., Raven Press, New York (1989), which is incorporated herein by reference.)

One important lymphokine is γ-IFN, which is secreted by some T cells. In addition to its anti-viral activity, γ-IFN stimulates natural killer (NK) cells, activates macrophages, and stimulates the expression of MHC molecules on the surface of cells (Paul, op. cit., pp. 622-624). Hence γ-IFN generally serves to enhance many aspects of immune function, and is a logical candidate for a therapeutic drug in cases where such enhancement is desired, e.g., in treating cancer. Conversely, in disease states where the immune system is over-active, e.g., autoimmune diseases and organ transplant rejection, antagonists of γ-IFN may be used to treat the disease by neutralizing the stimulatory effects of γ-IFN.

One class of effective antagonists of γ-IFN are monoclonal antibodies that bind to and neutralize it (see, e.g., Van der Meide et al., J. Gen. Virol, 67, 1059 (1986)). In in vitro and in vivo mouse models of transplants, anti-γ-IFN antibodies have been shown to delay or prevent rejection (Landolfo et al., Science 229, 176 (1985) and Rosenberg et al., J. Immunol. 144, 4648 (1990), both of which are incorporated herein by reference). Treatment of mice prone to develop a syndrome like systemic lupus erythematosus (SLE) with a monoclonal antibody to γ-IFN significantly delayed onset of the disease (Jacob et al., J. Exp. Med. 166, 798 (1987)). Under some conditions, an anti-γ-IFN antibody alleviated adjuvant arthritis in rats (Jacob et al., J. Immunol. 142, 1500 (1989)), suggesting that anti-γ-IFN may be effective against some cases of rheumatoid arthritis in human patients. Multiple sclerosis (MS) in patients is made worse by treatment with γ-IFN (Panitch et al., Neurology 36 (suppl. 1), 285 (1986)), so an anti-γ-IFN antibody may alleviate MS. Thus, an anti-γ-IFN antibody may be effective in treating these and other autoimmune diseases.

For treatment of human patients, a murine monoclonal that binds to and neutralizes human γ-IFN (see, e.g., Yamamoto et al., Microbiol. Immunol. 32, 339 (1988)) may be used. Another murine monoclonal antibody designated AF2 that neutralizes human γ-IFN, and inhibits binding of γ-IFN to its cellular receptor, is disclosed herein. Unfortunately, the use of non-human monoclonal antibodies such as AF2 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, have a relatively short circulating half-life in humans, and lack other important immunoglobulin functional

characteristics when used in humans.

The present invention provides novel compositions useful, for example, in the treatment of human autoimmune disorders, the compositions containing humanized immunoglobulins specifically capable of binding to γ-IFN. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to γ-IFN at affinity levels stronger than about 10^7 M^{-1}. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to γ-IFN.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such as a non-steroidal anti-inflammatory drug, a corticosteroid, or an immunosuppressant. All of these compounds will be particularly useful in treating autoimmune disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with γ-IFN epitopes are provided. These immunoglobulins, which have binding affinities to γ-IFN of at least about 10^7 M^{-1}, and preferably 10^8 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., neutralizing human γ-IFN. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with γ-IFN. In a preferred embodiment, one or more of the CDR's will come from the AF2 antibody. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of autoimmune disorders in human patients by a variety of techniques.

The antibodies of the present invention will typically find use individually in treating autoimmune conditions. For example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis,

systemic lupus erythematosus, and myasthenia gravis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with other lymphokines or lymphokine receptors. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include interleukins 1 through 10 and the p55 and p75 chains of the IL-2 receptor (see, Waldmann, Annu. Rev. Biochem. 58, 875 (1989) and Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), both of which are incorporated herein by reference). Other antigens include those on cells responsible for the disease, e.g., the so-called "Clusters of Differentiation" (Leucocyte Typing III, ed. by A.J. McMichael, Oxford University Press (1987), which is incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include non-steroidal anti-inflammatory agents (e.g., aspirin, ibuprofen), steroids (e.g., prednisone) and immunosuppressants (e.g., cyclosporin A, cytoxan), but numerous additional agents well-known to those skilled in the art may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins, e.g., to kill γ-IFN-secreting cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a γ-IFN epitope.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of γ-IFN antigens, or the like.

The following examples are offered by way of illustration, not by limitation.

EXPERIMENTAL

Example 1: Humanized anti-Tac antibody

Design of genes for humanized anti-Tac light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, E. Kabat et al., U.S. Dept. of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain variable region of anti-Tac is more homologous to the heavy chain of this antibody than to any other complete heavy chain variable region sequence in the National Biomedical Foundation Protein Identification Resource.

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence (Fig. 1B, upper lines; see, commonly assigned U.S. Serial Numbers 07/223,037 filed 09/28/88, and 07/181,862 filed 04/15/88, both of which are now abandoned and which are incorporated herein by reference) was aligned with the sequence of the Eu heavy chain (Fig. 1B,

lower lines). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of four categories defined above, in which case the anti-Tac amino acid was selected:

(1) The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31-35, 50-66, 99-106);

(2) The Eu amino acid was rare for human heavy chains at that position, whereas the anti-Tac amino acid was common for human heavy chains at that position (amino acids 27, 93, 95, 98, 107-109, 111);

(3) The position was immediately adjacent to a CDR in the amino acid sequence of the anti-Tac heavy chain (amino acids 30 and 67); or

(4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen binding region (amino acids 48 and 68).

Amino acid #27 is listed in category (2) because the acceptor Eu amino acid Gly is rare, and the donor anti-Tac amino acid Tyr is chemically similar to the amino acid Phe, which is common, but the substitution was actually made because #27 also fell in category (4). Although some amino acids fell in more than one of these categories, they are only listed in one. The amino acids in the humanized heavy and light chains are numbered according to the lower lines of Fig. 1A and Fig. 1B.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (Fig. 1A, lower lines). The Eu amino acid was selected at each position for the humanized sequence, unless the position again fell into one of the categories (1) - (4):

(1) CDR's (amino acids 24-34, 50-56, 89-97);

(2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63);

(3) Adjacent to CDR's (no amino acids; Eu and anti-Tac were already the same at all these positions); or

(4) Possible 3-dimensional proximity to binding region (amino acid 60).

The actual nucleotide sequence of the heavy and light chain genes were selected as follows:

(1) The nucleotide sequences code for the amino acid sequences chosen as described above;

(2) 5' of these coding sequences, the nucleotide sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies;

(3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals; and

(4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain genes

To synthesize the heavy chain, four oligonucleotides were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing.

Together, the oligonucleotides cover the entire humanized heavy chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 µl of TA (33 mM Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 µM each, heated to 95°C for 4 min. and cooled slowly to 4°C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide, the following components were added in a final volume of 100 µl:

10 µl	annealed oligonucleotides
0.16 mM each	deoxyribonucleotide
0.5 mM	ATP
0.5 mM	DTT
100 µg/ml	BSA
3.5 µg/ml	T4 g43 protein (DNA polymerase)
25 µg/ml	T4 g44/62 protein (polymerase accessory protein)
25 µg/ml	45 protein (polymerase accessory protein)

The mixture was incubated at 37°C for 30 min. Then 10 u of T4 DNA ligase was added and incubation at 37°C resumed for 30 min. The polymerase and ligase were inactivated by incubation of the reaction at 70°C for 15 min. To digest the gene with Xba I, to the reaction was added 50 µl of 2x TA containing BSA at 200 µg/ml and DTT at 1 mM, 43 µl of water, and 50 u of Xba I in 5 µl. The reaction was incubated for 3 hr at 37°C, and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUC19 by standard methods. Four plasmid isolates were purified and sequenced using the dideoxy method. One of these had the correct sequence.

To synthesize the light chain, four oligonucleotides JFD1, JFD2, JFD3, JFD4 were synthesized. Two of the oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing. Together, the oligonucleotides

cover the entire humanized light chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 µg each of JFD1 and JFD2 were combined in 20 µl sequence buffer (40 mM Tris-HCl, pH 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70°C for 3 min and allowed to cool slowly to 23°C in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 6.5 u of sequenase (US Biochemicals) was added, in a final volume of 24 µl, and incubated for 1 hr at 37°C to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 overlap and therefore in each of the synthesized DNAs). The reactions were run on polyacrylamide gels, and the Xba I - Hind III fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and correct ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then inserted into the Xba I site of the vector pVγ1 (see, commonly assigned U.S. Serial Number 07/223,037 filed 09/28/88, now abandoned, which is incorporated herein by reference) in the correct orientation by standard methods, to produce the plasmid pHuGTAC1. This plasmid will express high levels of a complete heavy chain when transfected into an appropriate host cell.

The two light chain Xba I - Hind III fragments were isolated from the pUC18 plasmids in which they had been inserted. The vector plasmid pVκ1 (see, commonly assigned U.S. Serial Number 07/223,037 filed 09/28/88, now abandoned, which is incorporated herein by reference) was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the circular form: vector - Xba I - fragment 1 - Hind III - fragment 2 - Xba I - vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC, therefore contains the complete humanized light chain and will express high levels of the light chain when transfected into an appropriate host cell.

Synthesis and affinity of humanized antibody

The plasmids pHuGTAC1 and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was incubated with HUT-102 cells that are known to express the IL-2 receptor. After washing, the cells were incubated with

fluorescein-conjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (Fig. 7A), clearly show that the humanized antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (Fig. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells, giving similar results.

For the next experiments, cells producing the humanized antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, CA) according to standard techniques. To determine the affinity of the humanized antibody relative to the original anti-Tac antibody, a competitive binding experiment was performed. About 5 x 10^5 HUT-102 cells were incubated with known quantities (10 - 40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4°C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4°C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess.

Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4°C with 250 ng of phycoerythrin- conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FACSCAN cytofluorometer.

Use of increasing amounts (10 - 40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrin-conjugated avidin that bound in the last step, thus decreasing fluorescence (Fig. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (Fig. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more.

Example 2: A second humanized anti-Tac antibody

Higher level expression of the humanized anti-Tac antibody

Three new plasmid vectors were prepared for expression of the humanized antibodies. The plasmid pVg1 (Fig. 9A) contains a human cytomegalovirus IE1 promoter and enhancer (Boshart et al., Cell 41, 521 (1985), which is incorporated herein by reference), the human genomic Cγ1 segment including part of the preceding intron, and the hygomycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 9B) is similar to pVg1 but contains the human genomic Cκ segment and the gpt gene. The

plasmid pVg1-dhfr was constructed similarly to pVg1 but contains a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) in place of the hygomycin gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and cloned into the Xba I sites of the plasmid vectors pVk and pVg1. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were then transfected with the plasmid encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture supernatant of transfected cells by protein A sepharose chromatography.

Construction of the second humanized anti-Tac antibody

To determine whether it was actually necessary to use the mouse anti-Tac amino acids in categories (2) - (4) in the humanized anti-Tac antibody to retain binding affinity, a second humanized anti-Tac antibody was constructed. In the second antibody, only mouse anti-Tac amino acids in Category (1), i.e., in the CDR's themselves, were used, with all other amino acids coming from the human Eu framework. For purposes of this discussion, the original humanized anti-Tac antibody will be called the "PDL humanized antibody," and the second humanized antibody will be called the "CDR-only humanized antibody." The amino acid sequences of the PDL and CDR-only humanized antibody (variable regions) are compared in Fig. 10A and Fig. 10B.

The CDR-only humanized anti-Tac heavy and light chain variable (V) region gene segments were constructed in essentially the same manner as the light chain of the PDL humanized anti-Tac immunoglobulin, as described above. Specifically, each V region gene segment was synthesized in two halves. For each half, two overlapping, opposite-strand oligonucleotides, approximately 110 to 130 bases in length (Fig. 11A and Fig. 11B), were annealed and extended with sequenase (U.S. Biochemicals). The resulting double strand fragments were digested with either Xba I and Hind III (light chain) or Xba I and Sal I (heavy chain) and inserted into plasmid pUC19. Clones with the correct sequence were identified by DNA sequencing. Complete heavy and light chain genes were generated by inserting the V region halves into the Xba I sites of pVg 1 and pVk respectively by three-fragment ligation.

The CDR-only humanized antibody was expressed in the same manner as the PDL humanized antibody, by transfecting first the light chain containing plasmid and then the heavy chain containing plasmid into SP2/0 cells. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody, which was purified by protein A sepharose chromatography. Antibody concentration was

determined by ELISA using purified PDL humanized antibody as a standard. That the purified CDR-only humanized antibody is assembled into H_2L_2 tetramers as expected was shown by analysis using reducing and non-reducing polyacrylamide gel electrophoresis.

The ability of the CDR-only humanized immunoglobulin to bind to the IL-2 receptor was assessed by fluorescence staining. Approximately 3.4 x 10^5 HUT-102 cells, which are known to highly express the IL-2 receptor on their surface, were incubated with 200 ng of either the PDL or CDR-only humanized antibody, washed, and then incubated with fluorescein-conjugated goat anti-human IgG antisera. Cell fluorescence was measured by flow cytometry with a FACScan (Becton Dickinson). As shown in Fig. 12, the PDL humanized antibody strongly stained the cells. However, staining by the CDR-only antibody was indistinguishable from staining by the negative control antibody humanized Fd79, which binds the gB glycoprotein of herpes simplex virus and not HUT-102 cells. Hence, by this assay, the CDR-only humanized antibody does not detectably bind the IL-2 receptor.

Binding of the PDL and CDR-only humanized anti-Tac antibodies to the IL-2 receptor were also compared in a competitive binding assay. Approximately 4 x 10^5 HUT-102 cells were incubated with 1.5 ng of radioiodinated mouse anti-Tac antibody (7 x 10^6 cpm/μg) and varying amounts of each humanized antibody (4 to 512 ng) in 200 μl total volume of binding buffer (RPMI 1040 medium, 10% fetal calf serum, 10 μg/ml murine IgG2a, 0.1% sodium azide). After incubation for 2 hours at 0°C, 800 μl of binding buffer was added, cells were collected by centrifugation and radioactivity was measured. The relative binding by the two humanized antibodies and by mouse anti-Tac is shown in a plot of bound/free labeled antibody versus competitor concentration (Fig. 13). The PDL humanized anti-Tac antibody affinity for IL-2 receptor is essentially equal to that of the mouse anti-Tac antibody, because it competes about equally well. But competition by the CDR-only humanized anti-Tac antibody to IL-2 receptor was undetectable at the antibody concentrations used, indicating a binding affinity reduction of at least 100-fold as compared to the PDL humanized anti-Tac antibody. Because the sequences of the PDL and CDR humanized anti-Tac antibodies differ only at positions where mouse framework residues in categories (2) - (4) were used in the PDL molecule, we conclude that at least one of these mouse framework residues are essential for high affinity binding.

Example 3: Construction of 5 other humanized antibodies

Cloning of heavy and light chain cDNAs

Five other humanized antibodies were designed and produced using the principles and categories disclosed herein. The antibodies are Fd79 and Fd138-80 which respectively bind to the gB and gD glycoproteins of herpes simplex virus (Metcalf et al., Intervirology 29, 39 (1988)), M195 (Tanimoto et al., Leukemia 3, 339 (1989)) which binds to the CD33 antigen, mik-β1 (Tusdo et al., Proc. Natl. Acad. Sci. USA 86, 1982 (1989)) which binds to the p75 chain of the IL-2 receptor, and CMV5 which binds to the gH glycoprotein of cytomegalovirus.

cDNAs for the heavy chain and light chain variable domain genes of each antibody were cloned using anchored polymerase chain reactions (Loh et al., Science 243, 219 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (Scheme shown in Fig. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For each antibody, at least two heavy chain and two kappa clones were sequenced and found to have the same sequence. The deduced amino acid sequences of the mature light and heavy chain variable regions are shown in Figs. 2A-6B, upper lines.

Design of humanized antibodies

In order to retain high binding affinity of the humanized antibodies, the principles and categories described above were utilized when designing the antibodies. Based on high sequence homology, human antibodies were selected to provide both the acceptor light and heavy chain human frameworks for the mouse antibodies, as follows: human Pom for Fd79, human Eu for Fd138-80, human Eu for M195, human Lay for mik-β1, and human Wol for CMV5.

The computer programs ABMOD and ENCAD (Levitt, J. Mol. Biol., 168, 595 (1983) and Zilber et al., Biochemistry 29, 10032 (1990), both of which are incorporated herein by reference) was used to construct a model of the variable region of each mouse antibody. The model was used to determine the amino acids in each framework that were close enough to the CDR's to potentially interact with them (category 4 above). For each antibody, the positions found to fall in the categories (1) - (5) defined above are given in Table 1, numbered as in Figs. 2A-6B.

TABLE 1

Fd79 Antibody

Category	Light Chain	Heavy Chain
1	24-38, 54-50, 93-100	31-35, 50-66, 99-111
2	9, 45, 46, 83	82, 112
3	53	112
4	53	97
5	81	

Fd138-80 Antibody

Category	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-110
2	48, 63	93, 98, 111, 112, 113, 115
3	--	30, 67, 98, 111
4	36, 48, 87	27, 30, 37, 48, 67, 68, 98

M195 Antibody

Category	Light Chain	Heavy Chain
1	24-38, 54-60, 93-101	31-35, 50-66, 95-105
2	10, 52, 67, 110	93, 95, 98, 106, 107 108, 110
3	--	30, 67, 98, 106
4	40, 52, 74	27, 30, 48, 68, 98

mik-β1 Antibody

Category	Light Chain	Heavy Chain
1	24-33, 49-55, 88-96	31-35, 50-65, 98-108
2	13	84, 89, 90
3	--	30, 49
4	70	29, 30, 72, 73
5	41	1

CMV5 Antibody

Category	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-108
2	--	69, 80
3	49	30
4	49	24, 27, 28, 30, 97
5	--	5

In designing each humanized antibody, at each position the amino acid was selected to be the same as in the human acceptor sequence, unless the position fell in categories (1) - (4), in which case the amino acid from the mouse donor sequence was used, or in category (5), in which case an amino acid typical for human sequences at that position was used.

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including signal peptides typically from the mouse antibody chains, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included splice donor signals typical for immunoglobulin genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 base long with a 15-20 base overlap. Double stranded DNA fragments were synthesized with Klenow or Taq polymerase or sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of pVg1 (heavy chains of Fd79 and Fd138-80) or pVg1-dhfr (heavy chains of M195, mik-β1, CMV5) or pVk (all light chains) expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.).

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2 M Glycine-HCl, pH 3.0 and neutralized with 1 M Tris pH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of the humanized antibodies

The binding of the humanized antibodies to cell types expressing the corresponding antigens was tested: HSV-infected cells for Fd79 and Fd138-80, U937 cells for M195, YTJB cells for mik-β1 and CMV-infected cells for CMV5. By fluorocytometry, the humanized antibodies bind approximately as well as the original mouse antibodies and the corresponding chimeric antibodies. Moreover, the humanized antibodies compete approximately as well as the

corresponding mouse antibodies against the radiolabeled mouse antibodies for binding to the cells, so the humanized antibodies have approximately the same binding affinity as the mouse antibodies, typically within about 2 fold or better, see, e.g., Table 2.

TABLE 2

Binding affinities of murine and humanized antibodies.

	Mouse K_a (M^{-1})	Humanized K_a (M^{-1})
Fd79 (anti-gB)	1.1×10^8	5.3×10^7
Fd138-80 (anti-gD)	5.2×10^7	4.8×10^7

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other antibodies. In comparison to other monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 4: Design of genes for anti-Tac human-like light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, Kabat, E., et al., U.S. Dept. of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain of anti-Tac is more homologous to the heavy chain of this antibody than to any other heavy chain sequence in the National Biomedical Foundation Protein Identification Resource.

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence was aligned with the sequence of the Eu heavy chain (Fig. 15). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of the following categories, in which case the anti-Tac amino acid was selected.

(1) The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31-35, 50-66, 99-106);

(2) The Eu amino acid was unusual for human heavy chains at that position, whereas the anti-Tac amino acid was typical for human heavy chains at that position (amino acids 27, 93, 95, 98, 107-109, 111);

(3) The position was immediately adjacent to a CDR in the amino

acid sequence of the anti-Tac heavy chain (amino acids 30 and 67).

(4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen binding region (amino acids 48 and 68).

Some amino acids fell in more than one of these categories but are only listed in one.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (Fig. 16). The Eu amino acid was selected at each position, unless the position again fell into one of the categories (1) - (4), (with light chain replacing heavy chain in the category definitions):

(1) CDRs (amino acids 24-34, 50-56, 89-97).
(2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63).
(3) Adjacent to CDRs (no amino acids; Eu and anti-Tac were already the same at all these positions).
(4) Possible 3-dimensional proximity to binding region (amino acid 60).

The actual nucleotide sequence of the heavy (Fig. 17) and light chain (Fig. 18) genes were selected as follows:

(1) the nucleotide sequences code for the amino acid sequences chosen as described above.
(2) 5' of these coding sequences, the nucleotide sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies.
(3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals.
(4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain genes

To synthesize the heavy chain, four oligonucleotides HES12, HES13, HES14, HES15 (Fig. 19A) were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (Fig. 19B). Together, the oligonucleotides cover the entire humanized heavy chain (Fig. 17) with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 µl of TA (33 mM Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 µM each, heated to 95°C for 4 min. and cooled slowly to 4°C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide (Fig. 19B), the following components were added in a final volume of 100 µl:

10 µl	annealed oligonucleotides
0.16 mM each	deoxyribonucleotide
0.5 mM	ATP
0.5 mM	DTT
100 µg/ml	BSA
3.5 µg/ml	T4 g43 protein (DNA polymerase)
25 µg/ml	T4 g44/62 protein (polymerase accessory protein)
25 µg/ml	45 protein (polymerase accessory protein)

The mixture was incubated at 37°C for 30 min. Then 10 U of T4 DNA ligase was added and incubation at 37°C resumed for 30 min. The polymerase and ligase were inactivated by incubation of the reaction at 70°C for 15 min. To digest the gene with Xba I, to the reaction was added 50 µl of 2x TA containing BSA at 200 µg/ml and DTT at 1 mM, 43 µl of water, and 50 U of Xba I in 5 µl. The reaction was incubated for 3 hr at 37°C, and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUC19 by standard methods. Four plasmid isolates were purified and sequenced using the dideoxy method. One of these had the correct sequence (Fig. 17).

To synthesize the light chain, four oligonucleotides JFD1, JFD2, JFD3, JFD4 (Fig. 20A) were synthesized. Two of the oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (Fig. 20B). Together, the oligonucleotides cover the entire humanized light chain (Fig. 18) with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 µg each of JFD1 and JFD2 were combined in 20 µl sequenase buffer (40 mM Tris-HCl, pH 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70°C for 3 min and allowed to cool slowly to 23°C in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 6.5 U of sequenase (US Biochemicals) was added, in a final volume of 24 µl, and incubated for 1 hr at 37°C to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to

each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 overlap and therefore in each of the synthesized DNAs; Fig. 20B). The reactions were run on polyacrylamide gels, and the Xba I - Hind III fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and correct ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then inserted into the Xba I site of the vector pVγ1 in the correct orientation by standard methods, to produce the plasmid pHuGTAC1 (Fig. 21). This plasmid will express high levels of a complete heavy chain when transfected into an appropriate host cell.

The two light chain Xba I - Hind III fragments were isolated from the pUC18 plasmids in which they had been inserted. The vector plasmid pVκ1 was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the circular form: vector - Xba I - fragment 1 - Hind III - fragment 2 - Xba I - vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC (Fig. 22), therefore contains the complete humanized light chain (Fig. 18) and will express high levels of the light chain when transfected into an appropriate host cell.

Synthesis and affinity of humanized antibody

The plasmids pHuGTAC1 and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids (Figs. 21, 22) by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was incubated with HUT-102 cells that are known to express the IL-2 receptor. After washing, the cells were incubated with fluorescein-conjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (Fig. 7A), clearly show that the humanized antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (Fig. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells (Fig. 7B and Fig. 7C), giving similar results.

For further experiments, cells producing the humanized antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, CA) according to standard techniques. To determine the affinity of the humanized antibody relative to the original anti-Tac antibody, a competitive binding experiment

was performed. About 5 x 10^5 HUT-102 cells were incubated with known quantities (10 - 40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4°C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4°C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess. Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4°C with 250 ng of phycoerythrin-conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FACSCAN cytofluorometer.

Use of increasing amounts (10 - 40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrin-conjugated avidin that bound in the last step, thus decreasing fluorescence (Fig. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (Fig. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more.

Biological properties of the humanized antibody

For optimal use in treatment of human disease, the humanized antibody should be able to destroy T-cells in the body that express the IL-2 receptor. One mechanism by which antibodies may destroy target cells is antibody-dependent cell-mediated cytotoxicity, abbreviated ADCC (Fundamental Immunology, Paul, W., Ed., Raven Press, New York (1984), at pg. 681), in which the antibody forms a bridge between the target cell and an effector cell such as a macrophage that can lyse the target. To determine whether the humanized antibody and the original mouse anti-Tac antibody can mediate ADCC, a chromium release assay was performed by standard methods. Specifically, human leukemia HUT-102 cells, which express the IL-2 receptor, were incubated with ^{51}Cr to allow them to absorb this radionuclide. The HUT-102 cells were then incubated with an excess of either anti-Tac or humanized anti-Tac antibody. The HUT-102 cells were next incubated for 4 hrs with either a 30:1 or 100:1 ratio of effector cells, which were normal purified human peripheral blood mononuclear cells that had been activated by incubation for about 20 hrs with human recombinant IL-2. Release of ^{51}Cr, which indicated lysis of the target HUT-102 cells, was measured and the background subtracted (Table 3). The results show that at either ratio of effector cells, anti-Tac did not lyse a significant number of the target cells (less than 5%), while the humanized antibody did

(more than 20%). Hence, the humanized antibody is likely to be more efficacious than the original mouse antibody in treating T-cell leukemia or other T-cell mediated diseases.

TABLE 3

Percent ^{51}Cr release after ADCC

	Effector: Target ratio	
	30:1	100:1
Antibody		
Anti-Tac	4%	< 1%
Humanized anti-Tac	24%	23%

Higher level expression of the humanized anti-Tac antibody

Two new plasmid vectors were prepared for expression of the humanized antibody. The plasmid pVg1 (Fig. 9A) contains a human cytomegalovirus IE1 promoter and enhancer (Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and the hygomycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 9B) is similar to pVg1 but contains the human genomic Cκ segment and the gpt gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and cloned into the Xba I sites of the plasmid vectors pVk and pVG1. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were then transfected with the plasmid encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture supernatant of transfected cells by protein A sepharose chromatography.

From the foregoing, it will be appreciated that the human-like immunoglobulins of the present invention offer numerous advantages of other human IL-2 receptor-specific antibodies. In comparison to anti-Tac mouse monoclonal antibodies, the present human-like immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 5: Design of genes for mik-β1 humanized light and heavy chains

To exert its biological effects, IL-2 interacts with a specific

high-affinity membrane receptor (Greene, W., et al., Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem. 58, 875 (1989), which is incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide or alpha chain, being about 55kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., Proc. Nat. Acad. Sci. USA, 83, 9694 (1986) and Sharon et al., Science 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. The antibody, mik-β1, binds to the p75 chain (Tsudo et al., Proc. Nat. Acad. Sci. USA 86, 1982 (1989), which is incorporated herein by reference).

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E.Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in Fig. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC19 vector for sequencing. For mik-β1, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in Fig. 23A and Fig. 23B.

Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (Fig. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 24B) is similar to pVg1-dhfr but contains the human genomic Cκ segment and the gpt gene. Derivatives of the mik-β1 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals

and XbaI sites (see, C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric mik-β1 antibody was shown to bind to YTJB cells, which express the p75 antigen, by flow cytometry (Fig. 25).

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence database (performed with the MicrorGenie Sequence Analysis Software (Beckman)), the antibody Lay was chosen to provide the framework sequences for humanization of mik-β1.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the mik-β1 variable region. The model was used to determine the amino acids in the mik-β1 framework that were close enough to the CDRs to potentially interact with them (category 4 below). To design the humanized light and heavy chain mik-β1 variable regions, at each position the amino acid was chosen to be the same as in the Lay antibody, unless that position fell in one or more of five categories:

(1) The position fell within a CDR,

(2) The Lay amino acid was unusual for human antibodies at that position, whereas the mik-β1 amino acid was typical for human antibodies at that position.

(3) The position was immediately adjacent to a CDR,

(4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDRs).

For positions in these categories, the amino acid from the (mouse) mik-β1 antibody was used. In addition, a position was in the fifth category if

(5) The Lay amino acid was highly unusual for human antibodies at that position, and the mik-β1 amino

acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 4. Some amino acids may be in more than one category. The final sequences of the humanized mik-β1 light and heavy chain variable domains are shown in Fig. 26A and Fig. 26B, compared with the Lay sequences.

TABLE 4

Category	Light Chain	Heavy Chain
1	24-33, 49-55, 88-96	31-35, 50-65, 98-108
2	13	84, 89, 90
3		30, 49
4	70	29, 30, 72, 73
5	41	1

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse mik-β1 chains (Fig. 23A and Fig. 23B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (Fig. 27A and Fig. 27B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 base long with about a 20 base overlap. Double stranded DNA fragments were synthesized with sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pBluescriptII KS (+) (Stratagene) vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors. In vitro mutagenesis was used to change an Ala amino acid originally encoded by oligonucleotide wps54 to the Glu (E) at position 1 of the humanized heavy chain (Fig. 26B) by changing the nucleotides CT to AG. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ElISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant

over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2 M Glycine-HC1, pH 3.0 and neutralized with 1 M Tris PH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized mik-β1 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized antibody bound to YTJB cells, which express p75 chain at a high level, in a fluorocytometric analysis in a manner similar to the chimeric antibody (Fig. 25), showing that it recognizes the same p75 protein.

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse mik-β1 antibody (Fig. 28). The binding affinities were calculated according to the methods of Berzofsky (J.A. Berzofsky and I.J. Berkower, in Fundamental Immunology (ed. W.E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The binding affinity of the humanized mik-β1 antibody was within about 2-fold of the affinity of the mouse mik-β1 antibody.

The ability of humanized mik-β1 plus humanized anti-Tac antibody to inhibit IL-2 stimulated proliferation of human lymphocytes was determined. Human mononuclear cells, collected from human blood by centrifugation on Ficoll-Paque (Pharmacia), were diluted to 2×10^6 cells/ml in RPMI medium + 10% fetal calf serum (FCS). A 1/200 volume of phytohemagglutinin P (Difco) was added and the cells were incubated for 4 days. The cells were incubated an additional 4 days in RPMI + 10% FCS + 10 u/ml IL-2. 10^5 of these PHA activated blasts were then incubated with or without 2 µg each of humanized mik-β1 and humanized anti-Tac in 150 µl of RPMI + 10% FCS in wells of a 96-well plate for 1 hr, to which various dilutions of IL-2 (Amgen) were then added in 50 µl medium. The cells were incubated 48 hr, 0.5 µCi methyl-^{3}H-thymidine (Amersham, 82 Ci/mmol) was added, and the cells were incubated 24 hr. Cells were harvested with a cell harvester and radioactivity determined. The combination of the antibodies greatly inhibited proliferation of the cells in response to IL-2 (Fig. 29), suggesting a combination of the antibodies will have strong immunosuppressive properties. Humanized mik-β1 plus humanized anti-Tac inhibited proliferation much more strongly than did either antibody alone.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other p75 specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 6: Design of genes for Fd79 and Fd138-80 humanized light and heavy

chains

Exemplary DNA sequences coding for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd79 and Fd138-80, are shown in Fig. 30A through Fig. 30D.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain regions (E.Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in Fig. 14). This method yields clones with authentic variable domain sequences, in contrast to other methods using mixed primers designed to anneal to the variable domain sequence (J.W. Larrick et al., Bio/Technology 7, 934 (1989) and Y.L. Chiang et al., BioTech. 7, 360 (1989)). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For Fd79, two gamma-1 specific and 5 kappa specific clones were sequenced. The two gamma-1 specific clones are identical in sequence. This heavy chain cDNA fragment encodes a signal peptide of 19 amino acids, a V region in mouse heavy chain subgroup IIIB, a D segment, and a J_H1 segment with 4 alterations compared to the genomic J_H1 sequence. The deduced amino acid sequence is shown in Fig. 30A.

The five kappa specific clones belong to two groups. Two clones are identical and encode a kappa chain in which the conserved amino acid 23 cysteine has been substituted by a tyrosine, probably representing the non-productive allele. The other three clones have an identical sequence encoding a signal peptide sequence of 20 amino acids, a V region in mouse kappa chain subgroup III, and a J_k2 segment with a single alteration compared to the genomic J_k2 sequence (Fig. 30B). The validity of the heavy chain and the kappa chain sequences was subsequently confirmed by the construction and expression of a chimeric antibody as discussed below.

The heavy chain and the kappa chain of Fd138-80 were cloned similarly. Three clones each of the heavy chain and the kappa chain were sequenced. All three heavy chain clones have an identical sequence encoding a signal peptide sequence of 19 amino acids, a V region in mouse heavy chain subgroup II, a D segment and the J_H3 segment (Fig. 30C). The three kappa clones are also identical in sequence. This light chain fragment encodes a signal peptide sequence of 20 amino acids, a V region gene in mouse kappa chain subgroup V and the J_k5 segment (Fig. 30D). Both chains shown no irregularities in coding sequence; their validity was subsequently confirmed by construction and expression of a chimeric antibody.

Construction and expression of chimeric antibodies.

Two plasmid vectors were prepared for construction and expression of

the chimeric antibody genes. The plasmid pVg1 (Fig. 9A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 9B) is similar to pVg1 but contains the human genomic Cκ segment and the gpt gene. Derivatives of the Fd79 and Fd138-80 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibodies, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric Fd79 and Fd138-80 antibodies were shown to bind to HSV-1 infected vero cells by flow cytometry. Viral neutralization assays also indicated that the chimeric antibodies retain the neutralization activities of the murine antibodies (data not shown, but see below for similar results with humanized antibodies).

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Pom was chosen to provide the framework sequences for humanization of Fd79.

The computer program ENCAD (Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the Fd79 variable region. Inspection of the refined model of murine Fd79 revealed two amino acid residues in the framework that are close enough to have significant contacts with the CDR residues (Table 5). Lys in light chain position 49 has contacts with 3 amino acids in CDR2 of the light chain (L50

Tyr, L53 Asn, L55 Glu) and 2 amino acids in CDR3 of the heavy chain (H99 Asp, H100 Tyr). Leu in heavy chain position 93 also shows interactions with 2 amino acids in CDR2 of the heavy chain (H35 Ser, H37 Val) and an amino acid in CDR3 of the heavy chain (H100C Phe). Hence, L49 Lys and H93 Leu were retained in the construction of humanized Fd79, as their replacement with human Pom framework residues would be likely to introduce distortions into the CDRs. Also, 7 other residues in the Pom framework (5 in the light chain and 2 in the heavy chain) were substituted with common human residues (identical to the murine Fd79 sequence in 6 of the choices) because of their rare occurrence in other human antibodies. The elimination of unusual amino acids in the framework may further reduce immunogenicity. The murine Fd79 sequences and the corresponding humanized sequences are shown in Fig. 30A and Fig. 30B. Substituted residues in the Pom framework are underlined.

TABLE 5

Residues in the framework sequence showing contacts with residues in the hypervariable regions.

Residue No.[1]	Amino Acid	Contacting CDR residues[2]
Fd79		
L49	Lys	L50Y, L53N, L55E, H99D, H100Y
H93	Leu	H35S, H37V, H100CF
Fd138-80		
L36	His	L34V, L89Q
H27	Tyr	H32H, H34I
H30	Tyr	H32H, H53R
H48	Phe	H63F
H66	Lys	H63F
H67	Ala	H63F

1. The amino acid residues are numbered according to the Kabat system (E.A. Kabat et al., Sequences of Proteins of Immunological Interest, National Institutes of Health, Bethesda, MD (1987)): the first letter (H or L) stands for the heavy chain or light chain. The following number is the residue number. The last letter is the amino acid one letter code.

2. The hypervariable regions are defined according to Kabat: Light chain CDR1: residue 24-34; CDR2: 50-56; CDR3: 89-97. Heavy chain CDR1: 31-35; CDR2: 50-65; CDR3: 95-102.

Similarly, the murine heavy chain and light chain sequences of Fd138-80 were subjected to sequence homology search against the NBRF protein sequence database. The sequences of the human antibody Eu were selected to provide the framework sequences for humanized Fd138-80. Inspection of a computer-generated model of Fd138-80 revealed 6 amino acid residues in the

framework that are close enough to have important contacts with CDR residues. The residues and their contacting counterparts are listed in Table 5; these murine residues were retained in the construction of humanized Fd138-80. Two other residues (L87 Phe and H37 Met) show significant contacts with L98 Phe, which is immediately adjacent to CDR3, so these two mouse residues were also retained. Eight amino acids in the Eu framework (2 in the light chain and 6 in the heavy chain) were substituted with the murine residues (which are also consistent with the human consensus residues) because of their rare occurrence in other human antibodies. The murine Fd138-80 sequences and the corresponding humanized sequences are shown in Fig. 30C and Fig. 30D. Substituted residues in the Eu framework are underlined.

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the signal peptides, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase, digested with restriction enzymes, ligated to pUC18 vector and sequenced. The two fragments with the correct sequences were then ligated into the XbaI sites of pVg1 or pVk expression vectors.

The synthetic genes were then cloned into the pVg1 and pVk expression vectors. For each humanized antibody constructed, the heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibodies were purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2 M Glycine-HCl, pH 3.0 and neutralized with 1 M Tris PH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

<u>Properties of humanized antibodies</u>.

The humanized Fd79 and Fd138-80 antibodies were characterized in comparison to the murine and chimeric antibodies. Both humanized antibodies bind to Vero cells infected with HSV-1 in a fluorocytometric analysis in a manner similar to the chimeric antibodies (Fig. 31A and Fig. 31B), suggesting

that they recognize their respective viral antigens. To more quantitatively assess the binding activity, radioiodinated murine antibodies were bound to virally infected cells and Scatchard analysis performed.

The affinities of the humanized antibodies were determined by competition with the iodinated antibodies. Vero cells infected with HSV-1 were used as source of gB and gD antigens. Increasing amounts of competitor antibody (mouse or humanized) were added to 1.5 ng of radioiodinated tracer mouse antibody (2 μCi/μg) and incubated with 4 x 10^5 infected Vero cells in 0.2 ml of binding buffer (PBS + 2% FCS + 0.1% azide) for 1 hr. at 4°C. Cells were washed and pelleted, and their radioactivities were measured. The concentrations of bound and free tracer antibody were calculated. The binding affinities were calculated according to the methods of Berzofsky (J.A. Berzofsky and I.J. Berkower, in Fundamental Immunology (ed. W.E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference).

The measurements indicate that there is no significant loss of binding affinities in the humanized antibodies (Table 6). Specifically, there is an approximately 2-fold decrease in affinity in humanized Fd79 compared to the murine Fd79 (Ka of 5.3 x 10^7 M^{-1} vs. 1.1 x 10^8 M^{-1}). The affinity of humanized Fd138-80 is comparable to that of the murine antibody (Ka of 4.8 x 10^7 M^{-1} vs 5.2 x 10^7 M^{-1}).

TABLE 6

Binding affinities of murine and humanized antibodies.

	Mouse K_a (M^{-1})	Humanized K_a (M^{-1})
Fd79 (anti-gB)	1.1 x 10^8	5.3 x 10^7
Fd138-80 (anti-gD)	5.2 x 10^7	4.8 x 10^7

Murine Fd79 and Fd138-80 have been shown to neutralize HSV-1 in vitro without complement (J. Koga et al., Virology 151, 385 (1986)), so the neutralizing activities of the humanized antibodies were compared with the mouse antibodies. Serial dilutions of equal quantities of murine and humanized antibodies were incubated with virus for 1 hr. before inoculation onto Vero cells. After 4 days, cells were stained with neutral red to visualize plaques. Results from these plaque reduction assays indicated that both humanized Fd79 and Fd138-80 neutralize virus as efficiently as their murine counterparts (Figs. 32A and B). Both humanized and murine Fd79 cause a 90% reduction of plaques at an antibody concentration of 10 nM (1.5 μg/ml). Similarly, humanized and murine Fd138-80 were able to cause a 90% plaque reduction at equivalent levels.

The antibodies were also investigated for their ability to protect cells from viral spread in tissue culture. Vero cells were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37°C before addition of 10 μg/ml antibody. After four days, cells were stained with an anti-gB antibody for detection of viral antigens on infected cells. Results indicated that both murine and humanized Fd79 at 10 μg/ml protected culture cells from infection (Fig. 33A). However, neither murine nor humanized Fd138-80 were able to protect cells against viral spread (Fig. 33B), despite their ability to neutralize virus before inoculation. Both gB and gD are thought to be associated with cell fusion and virus infectivity (W. Cai et al., J. Virol. 62, 2596 (1988) and A.O. Fuller and P.G. Spear. Proc. Natl. Acad. Sci. USA 84, 5454 (1987)). However, it is possible that Fd79 blocks both the infectivity and cell fusion functions of gB, while Fd138-80 blocks only the infectivity function of gD, so virus can still spread cell-to-cell.

The binding, neutralization and protection results all indicate that the humanized Fd79 and Fd138-80 antibodies have retained the binding activities and the biological properties of the murine monoclonal antibodies. The availability of humanized antibodies with specificity for HSV gB and gD, inter alia, provides an opportunity for studies of the in vivo potency and immunogenicity of humanized antibodies in treating viral diseases. The recognition by Fd79 and Fd138-80 of type-common epitopes of gB and gD (J. Koga et al., Virology 151, 385 (1986)) expands the therapeutic potential to herpes simplex virus type 2 as well as type 1.

The use of a combination of two or more humanized antibodies in therapy is important for reducing the development of antibody resistant strains. Combination therapy of humanized antibodies with other antiviral agents such as acyclovir provides further opportunities to combat diseases when chemotherapeutic agents alone have not been effective. As Fd79 and Fd138-80 reduce the frequency of viral persistence in a murine ocular model (J.F. Metcalf et al., Cur. Eye Res. 6, 173 (1987)), the humanized antibodies, typically together with other antiviral agents, are capable of reducing episodes of recurrent genital infection, an area where traditional anti-viral agents have not been effective (L. Corey et al., N. Engl. J. Med. 306, 1313 (1982)). Incorporation of the human constant domains can also enhance effector functions, such as antibody-dependent cellular cytotoxicity, leading to greater therapeutic efficiency in human patients.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other HSV specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 7: Design of genes for M195 humanized light and heavy chains

The p67 protein or CD33 antigen is found on the surface of progenitors of myeloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or non-hematopoietic cells (see, Leucocyte Typing III, ed. by A.J. McMichael, Oxford University Press, pp. 622-629 (1987), which is incorporated herein by reference). Antibodies that are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

Another antibody that binds to CD33 is M195 (Tanimoto et al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference).

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E.Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in Fig. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For M195, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in Fig. 34A and Fig. 34B.

Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (Fig. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 24B) is similar to pVg1-dhfr but contains the human genomic Cκ segment and the gpt gene. Derivatives of the M195 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa

chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric M195 antibody was shown to bind to U937 cells, which express the CD33 antigen, by flow cytometry (Fig. 35).

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of M195.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the M195 variable region. The model was used to determine the amino acids in the M195 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain M195 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of four categories:

(1) The position fell within a CDR,
(2) The Eu amino acid was unusual for human antibodies at that position, whereas the M195 amino acid was typical for human antibodies at that position,
(3) The position was immediately adjacent to a CDR,
(4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse M195 antibody was used: The amino acids in each category are shown in Table 7. Some amino acids may be in more than one category. The final sequences of the humanized M195 light and heavy chain variable domains are shown in Fig. 36A and Fig.

36B, compared with the Eu sequences.

TABLE 7

Category	Light Chain	Heavy Chain
1	24-38, 54-60, 93-101	31-35, 50-66, 99-105
2	10, 52, 67, 110	93, 95, 98, 106, 107, 108, 110
3	--	30, 67, 98, 106
4	40, 52, 74	27, 30, 48, 68, 98

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse M195 chains (Fig. 34A and Fig. 34B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (Fig. 37A and Fig. 37B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2 M Glycine-HCl, pH 3.0 and neutralized with 1 M Tris PH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized M195 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized antibody bound to U937 cells in a fluorocytometric analysis in a manner similar to the chimeric antibody (Fig. 35), showing that it recognizes the same CD33 antigen.

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse M195 antibody (Fig. 38). The binding affinities were calculated according to the methods of Berzofsky (J.A. Berzofsky and I.J. Berkower, in Fundamental Immunology (ed. W.E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The mouse M195 had an affinity comparable to the published value (Tanimoto et al., op. cit.) and the humanized M195 antibody had an affinity the same as the mouse M195 to within experimental error.

Humanized M195 is useful to mediate antibody-dependent cellular cytotoxicity when human effector cells and human CD33-expressing cells are used. This is analogous to other humanized antibodies, such as reported by Junghans et al., Cancer Research 50, 1495 (1990), which is incorporated herein by reference.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CD33 specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 8: Design of genes for CMV5 humanized light and heavy chains

Three neutralizing antibodies to the gH glycoprotein of human cytomegalovirus (CMV) are designated CMV5, CMV109 and CMV115.

Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody CMV5 are included in Fig. 39A and Fig. 39B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVg1-dhfr (Fig. 24B and 24A) using site-directed mutagenesis, such as after CH1 to produce Fab fragments or after the hinge region to produce $(Fab')_2$ fragments.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E.Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in Fig. 14). The PCR amplified fragments were digested with EcoR I and HindIII and cloned into the pUC18 vector for sequencing. For CMV5, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in Fig. 39A and Fig. 39B. Similarly, by using techniques, which are well-known in the art, cDNAs for the CMV109 and CMV115 antibodies may be obtained and their sequence determined.

Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (Fig. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 24B) is similar to pVg1-dhfr but contains the human genomic Cκ segment and the gpt gene. Derivatives of the CMV5 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the cytomegalovirus promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric CMV5 antibody was shown to bind to CMV-infected cells, which express the gH antigen, by immunostaining of CMV-infected human embryonic lung fibroblasts.

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's

with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Wol was chosen to provide the framework sequences for humanization of CMV5.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the CMV5 variable region. The model was used to determine the amino acids in the CMV5 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain CMV5 variable regions, at each position the amino acid was chosen to be the same as in the Wol antibody, unless that position fell in one or more of five categories:

(1) The position fell within a CDR,

(2) The Wol amino acid was unusual for human antibodies at that position, whereas the CMV5 amino acid was typical for human antibodies at that position,

(3) The position was immediately adjacent to a CDR,

(4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Wol light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse CMV5 antibody was used. In addition, a position was in the fifth category if the Wol amino acid was highly unusual for human antibodies at that position, and the CMV5 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 8. Some amino acids may be in more than one category. The final sequences of the humanized CMV5 light and heavy chain variable domains are shown in Fig. 40A and Fig. 40B, compared with the Wol sequences.

TABLE 8

Category	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-108 69, 80
2		69, 80
3	49	30
4		24, 27, 28, 30, 97
5		5

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse CMV5 chains (Fig. 39A and Fig. 39B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (Fig. 41A and Fig. 41B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells are selected for gpt expression. Clones are screened by assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody is eluted with 0.2 M Glycine-HCl, pH 3.0 and neutralized with 1 M Tris PH 8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Humanized antibody was also produced by transient transfection. The heavy chain and light chain plasmids were transfected into S194 cells (ATCC TIB 19) by the DEAE-dextran method (Queen et al., Mol. Cell. Biol. 4, 1043

(1984), which is incorporated herein by reference), and humanized CMV5 antibody was purified from the media supernatant as above. Antibody was quantitated by ELISA assay for human Ig.

Properties of humanized antibodies.

The humanized CMV5 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized CMV5 antibody was shown to bind about as well as the mouse and chimeric antibodies to CMV antigen, by immunostaining of CMV-infected human embryonic lung (HEL) cells (ATCC CCL 137). HEL cells monolayers in 96-well plates were infected with CMV at 0.01 pfu/cell, incubated for 4 days, dried at 37°C and stored wrapped at 4°C. 100 µl blotto (5% Carnation Instant Milk in PBS at pH 7.4) was added to each well and incubated at 37°C for 30 min. The blotto was poured off and 75 µl of a series of 2-fold dilutions of mouse, chimeric and humanized CMV5 antibody was added to the wells. The plate was incubated 1 hr at 37°C and washed twice with blotto (each wash was left on for 10 min). Then 75 µl of diluted peroxidase (HRP) conjugated goat anti-mouse or anti-human IgG (Tago) was added to each well and incubated for 1 hr at 37°C. The plate was washed 2x with PBS and 150 µl of HRP substrate solution was added to each well. Color was allowed to develop at room temperature. The plates were washed with water and air dried. The wells were examined under a microscope to determine the highest dilution of the antibodies that formed a colored precipitate on the CMV-infected cells. For all three antibodies, 63 ng/ml was the least amount of antibody that produced a detectable precipitate, indicating that humanized CMV5 binds about as well as the mouse and chimeric antibodies.

To compare the affinities of mouse and humanized CMV5 in another way, a competition experiment was performed. Plates of CMV-infected HEL cells as above were incubated with blotto for 30 min at 37°C. The blotto was poured off and dilutions of mouse or humanized CMV5 were added to each well in 75 µl of PBS. Then 125 µl of radio-iodinated mouse CMV5 (1 µCi/µg) in PBS, containing 28,000 cpm was added to each well and incubated at 37°C for 2.5 hr. The plate was washed 5 times with PBS, and the contents of each well were solubilized with 200 µl of 2% SDS and counted. Increasing concentrations of mouse and humanized CMV5 inhibited binding of the radiolabeled CMV5 about equally well (Fig. 42), so humanized CMV5 has approximately the same binding affinity as mouse CV5. An irrelevant antibody did not compete in this assay.

The ability of humanized CMV5 to neutralize CMV is compared to that of mouse CMV5. Mouse and humanized CMV5 are successively diluted by 2-fold in 100 µl of DME medium + 2% FCS in wells of a 96-well plate. 100 µl of CMV, which has been diluted to contain 100 tissue culture infectious dose-50% (TCID50) units, are added to each well and incubated for 60 min at 37°C. Each well of antibody-virus mixture is added to a well of subconfluent HEL cells in

a 96-well plate from which the medium has been removed. The cells are incubated for 5 days and cytopathic effect (CPE) is examined in each well under a microscope. The highest dilution of antibody that inhibits CPE by 90% is a measure of the neutralizing ability of the antibody. The humanized CMV5 antibody will neutralize CMV antibody approximately as well as the mouse CMV5 antibody.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CMV specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Example 9: Design of genes for AF2 human-like light and heavy chains

This example is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of γ-IFN, such as monoclonal antibody AF2. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody AF2 are included in Fig. 43A and Fig. 43B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E.Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in Fig. 14). The PCR amplified fragments were digested with EcoR I and HindIII and cloned into the pUC18 vector for sequencing. For AF2, two gamma-2b specific and two kappa specific clones were sequenced. The two gamma-2b clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in Fig. 43A and Fig. 43B.

Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1-dhfr (Fig. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγ1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (Fig. 24B) is similar to pVg1-dhfr

but contains the human genomic Cκ segment and the gpt gene. Derivatives of the AF2 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Chimeric AF2 antibody was shown to bind to human γ-IFN by ELISA.

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of AF2.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the AF2 variable region. The model was used to determine the amino acids in the AF2 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain AF2 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of five categories:

(1) The position fell within a CDR,

(2) The Eu amino acid was unusual for human antibodies at that position, whereas the AF2 amino acid was typical for human antibodies at that position,

(3) The position was immediately adjacent to a CDR,

(4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse AF2 antibody was used. In addition, a position was in the fifth category if the Eu amino acid was highly unusual for human antibodies at that position, and the AF2 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 9. Some amino acids may be in more than one category. The final sequences of the humanized AF2 light and heavy chain variable domains are shown in Fig. 44A and Fig. 44B, compared with the Eu sequences.

TABLE 9

Category	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-106
2	48	93, 95, 98, 107, 108, 109, 111
3		30, 98, 107
4	48, 70	27, 28, 30, 98, 107
5	63	

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, plus typical immunoglobulin signal sequences, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (Fig. 45A and Fig. 45B) The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences are then ligated into the XbaI sites of

the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions are carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones are screened by assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody is eluted with 0.2 M Glycine-HCl, pH 3.0 and neutralized with 1 M Tris PH 8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized AF2 antibody is characterized in comparison to the murine and chimeric antibodies. The humanized antibody will bind to γ-IFN in an ELISA assay in a manner similar to the mouse and chimeric antibodies, showing that it recognizes γ-IFN.

To compare the binding affinities of mouse AF2 antibody and humanized AF2 antibody, a competitive ELISA assay is performed. An ELISA plate is coated with human recombinant γ-IFN by adding 100 μl of a 500 ng/ml solution of γ-IFN in PBS to each well and incubating overnight at 4°C. Subsequent steps are carried out at room temperature. The γ-IFN solution is removed and 200 μl of ELISA buffer (0.1% TWEEN-20, 1% Bovine serum albumin in PBS) is added to each well and incubated for 1 hr. After removing the solution, varying amounts of competitor antibody (mouse AF2 or humanized AF2) in 100 μl PBS is added to each well, along with an amount of biotinylated AF2 predetermined to give a good ELISA response. The plate is incubated for 1 hr and then washed 3 times with ELISA buffer. An amount of horseradish peroxidase (HRP)-conjugated streptoavidin predetermined to be in excess is added in 100 μl PBS to each well and incubated for 30 min. The plate is washed 3 times in ELISA buffer, and 100 μl of substrate solution for HRP is added to each well. The plate is incubated for 10-30 min, and the optical density of each well is determined with an ELISA reader (BioRad). The decrease in optical density with increasing concentrations of competitor antibodies mouse AF2 and humanized AF2 are plotted. Mouse AF2 and humanized AF2 will compete similarly, showing that their binding affinities for γ-IFN are approximately the same. The procedures used are well known in the art (e.g., Harlow and Lane, op. cit.).

An important biological activity of γ-IFN is the induction of expression of class II HLA antigens on cells. To determine the ability of mouse and humanized AF2 to neutralize this activity, about 5 x 10^4 HS294T cells (Basham et al., J. Immunol. 130, 1492 (1983), which is incorporated herein by reference) are plated in 1.0 ml DMEM medium + 10% FCS in each well of a 24-

well plate. After overnight incubation, 0.1 nM interferon and varying amounts of mouse or humanized AF2 are added to the cells, and the plate is incubated for 72 hr. The cells are removed from the plate with 0.05 M EDTA, stained with monoclonal antibody L243 from the American Type Culture Collection (ATCC) against HLA-D antigen, washed, stained with FITC conjugated goat anti-mouse Ig and analyzed with a FACScan (Becton-Dickinson). Increasing concentrations of mouse AF2 reduce fluorescence of the cells (Fig. 46), indicating the antibody is preventing induction of HLA-D by γ-IFN. The humanized AF2 will act similarly to mouse AF2 in this assay, showing that it neutralizes the biological activity of γ-IFN.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other γ-IFN specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be apparent that certain changes and modifications may be practiced within the scope of the appended claims.